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(54) Title: CHEMICALLY REGULATABLE AND ANTI-PATHOGENIC DNA SEQUENCES AND USES THEREOF (57) Abstract <p>The present invention provides recombinant or chimaeric DNA molecules comprising plant SAR genes, wherein the wild-type genes corresponding to said SAR genes can be chemically induced in a plant in a protein-synthesis independent or in a protein-synthesis dependent manner. Methods for obtaining these DNA molecules are also provided. Additionally, the present invention provides chemically inducible wheat genes, Arabidopsis chitinase IV, Maize PR-1mz, and Maize thaumatin PR-5mz. The anti-pathogenic sequences according to the invention can be genetically engineered and transformed into plants to obtain transgenic plants with enhanced resistance to disease. In one embodiment of the present invention, the chemically regulatable DNA promoter sequence of the Arabidopsis Pr-1 gene is provided. Another aspect of the present invention is a method of improving protection of a plant against a pest comprising transgenically expressing in said plant two or more DNA molecules encoding anti-pathogenic proteins, wherein the transgenically expressed proteins exert a synergistic effect.</p>		

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CHEMICALLY REGULATABLE AND ANTI-PATHOGENIC DNA SEQUENCES
AND USES THEREOF

The present invention relates to DNA molecules encoding proteins capable of conferring plant disease and/or plant pest resistance. Another aspect of the invention relates to the chemical regulation of gene expression. In particular, this invention relates to non-coding DNA sequences which, in the presence of chemical regulators, regulate the transcription of associated DNA sequences in plants. Both aspects of the invention relate, in part, to genes associated with the response of plants to pathogens.

Advances in recombinant DNA technology coupled with advances in plant transformation and regeneration technology have made it possible to introduce new genetic material into plant cells, plants or plant tissue, thus introducing new traits, e.g., phenotypes, that enhance the value of the plant or plant tissue. Recent demonstrations of genetically engineered plants resistant to pathogens (EP-A 240 332 and EP-A 223 452) or insects (Vaeck, M. *et al.*, *Nature* 328: 33 (1987)) and the production of herbicide tolerant plants (DeBlock, M. *et al.*, *EMBO J.* 6: 2513 (1987)) highlight the potential for crop improvement. The target crops can range from trees and shrubs to ornamental flowers and field crops. Indeed, it is clear that the "crop" can also be a culture of plant tissue grown in a bioreactor as a source for some natural product.

Various methods are known in the art to accomplish the genetic transformation of plants and plant tissues (i.e., the stable introduction of foreign DNA into plants). These include transformation by Agrobacterium species and transformation by direct gene transfer.

A. tumefaciens is the etiologic agent of crown gall, a disease of a wide range of dicotyledons and gymnosperms, that results in the formation of tumors or galls in plant tissue at the site of infection. Agrobacterium, which normally infects the plant at wound sites, carries a large extrachromosomal element called the Ti (tumor-inducing) plasmid.

Ti plasmids contain two regions required for tumorigenicity. One region is the T-DNA (transferred-DNA) which is the DNA sequence that is ultimately found stably transferred to plant genomic DNA. The other region required for tumorigenicity is the vir (virulence) region which has been implicated in the transfer mechanism. Although the vir region is absolutely required for stable transformation, the vir DNA is not actually transferred to the infected plant. Transformation of plant cells mediated by infection with Agrobacterium tumefaciens

and subsequent transfer of the T-DNA alone have been well documented. See, for example, Bevan, M.W. and Chilton, M-D., *Int. Rev. Genet.* 16:357 (1982).

After several years of intense research in many laboratories, the Agrobacterium system has been developed to permit routine transformation of a variety of plant tissue. Representative species transformed in this manner include tobacco, tomato, sunflower, cotton, rapeseed, potato, soybean, and poplar. While the host range for Ti plasmid transformation using A. tumefaciens as the infecting agent is known to be very large, tobacco has been a host of choice in laboratory experiments because of its ease of manipulation.

Agrobacterium rhizogenes has also been used as a vector for plant transformation. This bacterium, which incites hairy root formation in many dicotyledonous plant species, carries a large extrachromosomal element called an Ri (root-inducing) plasmid which functions in a manner analogous to the Ti plasmid of A. tumefaciens. Transformation using A. rhizogenes has developed analogously to that of A. tumefaciens and has been successfully utilized to transform, for example, alfalfa, Solanum nigrum L., and poplar.

Several so-called direct gene transfer procedures have been developed to transform plants and plant tissues without the use of an Agrobacterium intermediate (see, for example, Koziel *et al.*, *Biotechnology* 11: 194-200 (1993) herein incorporated by reference). In the direct transformation of protoplasts the uptake of exogenous genetic material into a protoplast may be enhanced by use of a chemical agent or electric field. The exogenous material may then be integrated into the nuclear genome. The early work was conducted in the dicot tobacco where it was shown that the foreign DNA was incorporated and transmitted to progeny plants, see e.g. Paszkowski, J. *et al.*, *EMBO J.* 3: 2717 (1984); and Potrykus, I. *et al.*, *Mol. Gen. Genet.* 199: 169 (1985).

Monocot protoplasts have also been transformed by this procedure in, for example, Triticum monococcum, Lolium multiflorum (Italian ryegrass), maize, and Black Mexican sweet corn.

Alternatively exogenous DNA can be introduced into cells or protoplasts by microinjection. A solution of plasmid DNA is injected directly into the cell with a finely pulled glass needle. In this manner alfalfa protoplasts have been transformed by a variety of plasmids, see e.g. Reich, T.J. *et al.*, *Bio/Technology* 4: 1001 (1986).

A more recently developed procedure for direct gene transfer involves bombardment of cells by microprojectiles carrying DNA, see Klein, T.M. *et al.*, *Nature* 327: 70 (1987). In this procedure tungsten particles coated with the exogenous DNA are accelerated toward the target cells, resulting in at least transient expression in the example reported (onion).

Just as there is a variety of methods for the transformation of plant tissue, there is a variety of methods for the regeneration of plants from plant tissue. The particular method of regeneration will depend on the starting plant tissue and the particular plant species to be regenerated. In recent years it has become possible to regenerate many species of plants from callus tissue derived from plant explants. The plants which can be regenerated from callus include monocots, such as corn, rice, barley, wheat and rye, and dicots, such as sunflower, soybean, cotton, rapeseed and tobacco.

Regeneration of plants from tissue transformed with A. tumefaciens has been demonstrated for several species of plants. These include sunflower, tomato, white clover, rapeseed, cotton, tobacco, and poplar. The regeneration of alfalfa from tissue transformed with A. rhizogenes has also been demonstrated. Plant regeneration from protoplasts is a particularly useful technique, see Evans, D.A. *et al.*, in: "Handbook of Plant Cell Culture", Vol. 1, MacMillan Publ. Co., 1983, p. 124. When a plant species can be regenerated from protoplasts, then direct gene transfer procedures can be utilized, and transformation is not dependent on the use of A. tumefaciens. Regeneration of plants from protoplasts has been demonstrated for rice, tobacco, rapeseed, potato, eggplant, cucumber, poplar, and corn.

Various plant tissues may be utilized for transformation with foreign DNA. For instance, cotyledon shoot cultures of tomato have been utilized for Agrobacterium mediated transformation and regeneration of plants (see European application EP-A 249432). Further examples include Brassica species (see WO 87/07299) and woody plant species, particularly poplar (see U.S. Patent No. 4,795,855, incorporated by reference herein in its entirety).

The technological advances in plant transformation and regeneration technology highlight the potential for crop improvement via genetic engineering. There have been reports of genetically engineered tobacco and tomato plants which are resistant to infections of, for example, tobacco mosaic virus (TMV) and resistant to different classes of herbicides. Insect resistance has been engineered in tobacco and tomato plants.

The potential for genetic engineering is not limited to field crops but includes improvements in ornamentals, forage crops and trees. A less obvious goal for plant biotechnology, which includes both genetic engineering and tissue culture applications, is the enhanced production of a vast array of plant-derived chemical compounds including flavors, fragrances, pigments, natural sweeteners, industrial feedstocks, antimicrobials and pharmaceuticals. In most instances these compounds belong to a rather broad metabolic group, collectively denoted as secondary products. Plants may produce such secondary products to ward off potential predators, attract pollinators, or combat infectious diseases.

Plant cell cultures can be established from an impressive array of plant species and may be propagated in a bioreactor. Typical plant species include most of those that produce secondary products of commercial interest. It has been clearly demonstrated in a number of agriculturally important crop plants that stable genetic variants arising from the tissue culture of plant somatic cells (somaclonal variation) can be induced and selected. Numerous advantages flow from plant tissue culture production of secondary compounds. These include (1) the possibility of increased purity of the resultant product, (2) the conversion of inexpensive precursors into expensive end products by biotransformation, and (3) the potential for feeding substrate analogs to the culture to create novel compounds.

Whether the target of genetic engineering of plants is a field crop, ornamental shrub, flower, tree or a tissue culture for use in a bioreactor, a principal advantage to be realized is the control of expression of the chimeric gene so that it is expressed only at the appropriate time and to the appropriate extent, and in some situations in particular parts of the plant. For example, in order to achieve a desirable phenotype the chimeric gene may need to be expressed at levels of 1% of the total protein or higher. This may well be the case for fungal resistance due to chimeric chitinase expression or insect resistance due to increased proteinase inhibitor expression. In these cases the energy expended to produce high levels of the foreign protein may result in a detriment to the plant whereas, if the gene were expressed only when desired, for instance when a fungal or insect infestation is imminent, the drain on energy, and therefore yield, could be reduced.

Alternatively, the phenotype expressed by the chimeric gene could result in adverse effects to the plant if expressed at inappropriate times during development. For example, if the chimeric gene product were a plant hormone that induced pod abscission, early expression could bring about abscission of the fruit from the plant before the seed had

matured, resulting in decreased yield. In this case it would be much more advantageous to induce the expression of this type of gene to a time when pod abscission is preferred, or least injurious to the plant.

For tissue in culture or in a bioreactor the untimely production of a secondary product could lead to a decrease in the growth rate of the culture resulting in a decrease in the yield of the product. Therefore, it would be advantageous to allow the culture to grow without expressing the secondary product and then induce the chimeric gene at an appropriate time to allow for an optimized expression of the desired product.

In view of considerations like these, as well as others, it is clear that control of the time, extent and/or site of expression of the chimeric gene in plants or plant tissues would be highly desirable. Control that could be exercised easily in a field, a greenhouse or a bioreactor would be of particular commercial value.

External manipulation of the expression of endogenous genes which contain chemically regulatable sequences by the application of a chemical regulator is already known from Ward, E. *et al.*, *Plant Cell* 3: 1085-1094 (1991); Williams *et al.*, *Bio/Technology* 10: 540-543 (1992); and Uknes, S. *et al.*, *Plant Cell* 5: 159-169 (1993). The control provided is somewhat limited, however, due to the responsiveness of such sequences to endogenous chemical metabolites and cell signals as well as externally applied chemical regulators. In one aspect of the invention, alterations are taught which block the responsiveness of these genes to endogenous signals while maintaining responsiveness to externally applied chemical regulators.

Pest infestation of crop plants causes considerable loss of yield throughout the world and most crops grown in the U.S. suffer infestation, particularly from insects. Major insect pests in the U.S. include the European Corn Borer (*Ostrinia nubilalis*) in maize, the cotton bollworm (*Heliothis zea*) and the pink bollworm (*Pectinophora gossypiella*) in cotton and the tobacco budworm (*Heliothis virescens*) in tobacco. Resistance to pests is difficult to achieve using conventional breeding programs and typically pests have been controlled using chemical pesticides.

Recent advances in molecular biology and plant transformation technology have demonstrated the possibility of expressing in transgenic plants genes encoding insecticidal proteins; this represents a novel approach in the production of crop plants resistant to

pests. Most notably, the expression of genes encoding the *Bacillus thuringiensis* endotoxin has been successful in a wide range of plant species, and the analysis of transgenic lines expressing such genes has been well documented (Vaech *et al.*, *Nature* 328: 33-37 (1987); Fischhoff *et al.*, *Biotechnology* 5: 807-813 (1987); Carozzi *et al.*, *Plant Mol. Biol.* 20: 539-548 (1992); Koziel *et al.*, *Biotechnology* 11: 194-200 (1993)). Other insecticidal genes have been used successfully in generating insect resistant transgenic plants.

One approach has been the overexpression of genes encoding insect enzyme inhibitors such as trypsin inhibitors or seed proteins with known insecticidal activity, such as lectins (Hilder *et al.*, *Nature* 330: 160-163 (1987)). Indeed, plants expressing both the cowpea trypsin inhibitor and pea lectin were shown to have additive effects in providing insect resistance (Boulter *et al.*, *Crop Protection* 9: 351-354 (1990)). In cases where pests are able to infest parts of the plant or tissues not readily accessible to conventional pesticides, a transgenic approach may be more successful than the use of conventional pesticides.

For example, the tobacco budworm *Heliothis* is well known to be difficult to control using pesticides because it burrows deep into the plant tissue. Additionally, some pests of roots, such as nematodes, are not readily controlled by foliar applications of pesticides. An advantage in the use of transgenic plants expressing insecticidal proteins is the controlled expression of the proteins in all desired tissues.

Chitinases catalyze the hydrolysis of chitin, a β -1,4-linked homopolymer of *N*-acetyl-D-glucosamine. Several different plant chitinases have been described and the cDNA sequences for some of these have been reported (Meins *et al.*, *Mol. Gen. Genet.* 232: 460-469 (1992)). Based on structural characteristics, three classes have been distinguished. Class I chitinases have two structural domains, a cysteine-rich amino-terminal hevein domain and a carboxyterminal catalytic domain separated from the former by a variable spacer. Class II chitinases lack the cysteine-rich hevein domain and all or part of the variable spacer, but retain the catalytic domain. Class III chitinases lack the hevein domain and contain a catalytic domain that shares no significant homology with that of the class I or class II enzymes.

Class I chitinase gene expression is induced by ethylene, whereas class II and class III chitinase gene expression is induced in the SAR response. The chitinase/lysozyme disclosed in European patent application EP-A 392 225 and the chitinase/lysozyme disclosed in SEQ ID Nos. 20 and 21 are class III chitinases. It is well known that the level of chitinase activity of plants increases dramatically after pathogen invasion (Mauch *et al.*,

Plant Physiol. 76: 607-611 (1984)) and this is presumably due to the host plant's attempts to degrade the chitin of the fungal cell wall. Furthermore, chitinase has been shown *in vitro* to inhibit fungal and insect growth, and in transgenic plants a bacterial chitinase has been shown to exhibit inhibitory effects towards numerous pathogens and pests including insects (Suslow & Jones WPO 90-231246; U.S. Pat. Nos. 4,940,840 and 4,751,081; herein incorporated by reference in their entirety).

For over 90 years, scientists and naturalists have observed that when plants survive pathogen infection they develop an increased resistance to subsequent infections. In 1933, a phenomenon termed "physiological acquired immunity" was described in an extensive literature review by Chester, K.S., *Quart. Rev. Biol.* 8: 275-324 (1933). At that time, scientists believed they were investigating a phenomenon analogous to the immune response in mammals. In retrospect, at least three different processes were being called "acquired immunity": viral cross protection, antagonism (or biocontrol), and what we now refer to as systemic acquired resistance (SAR).

The first systematic study of SAR was published by A. Frank Ross in 1961. Using tobacco mosaic virus (TMV) on local lesion hosts, Ross demonstrated that infections of TMV were restricted by a prior infection. This resistance was effective against not only TMV, but also tobacco necrosis virus and certain bacterial pathogens. Ross coined the term "systemic acquired resistance" to refer to the inducible systemic resistance (Ross, A.F., *Virology* 14: 340-358 (1961)) and "localized acquired resistance" (LAR) to describe the resistance induced in inoculated leaves (Ross, A.F., *Virology* 14: 329-339 (1961)). It is still unclear whether SAR and LAR are aspects of the same response or distinct processes.

In the past 30 years, SAR has been demonstrated in many plant species and the spectrum of resistance has been broadened to include not only viruses and bacteria, but also many agronomically important phytopathogenic fungi (see Kuc, J., *BioScience* 32: 854-860 (1982)). However, understanding of the biochemical events leading to the establishment of SAR had not progressed substantially until the past dozen years. In 1982, the accumulation of a group of extracellular proteins called pathogenesis-related (PR) proteins were reported to correlate with the onset of SAR (Van Loon, L.C. *et al.*, *Neth. J. Plant. Path.* 88: 237-256 (1982)). In 1979, salicylic acid (SA) and certain benzoic acid derivatives were reported to be able to induce both resistance and the accumulation of PR proteins (White,

R.F., *Virology* 99: 410-412 (1979). As a result, SA was considered as a possible endogenous signal molecule (Van Loon, L.C. *et al.*, *Neth. J. Plant. Path.* 88: 237-256 (1982)). However, progress slowed through the 1980's and the involvement of PR proteins and salicylic acid in SAR was questioned.

With the advent of genetic engineering and recombinant DNA technology, the possibility of manipulating genetic material to improve the phenotype of plants has arisen. The present invention is based in part upon the discovery of the identity and role of genes involved in SAR which has allowed the application of modern molecular biological techniques for improved plant disease and plant pest resistance.

There are two major aspects of the present invention. The first aspect relates to chemically regulatable DNA sequences and the chemicals which regulate them. The second aspect relates to plant pathogenesis-related proteins. Both aspects of the invention have arisen, in part, from the inventors' identification and characterization of cDNAs and corresponding genes involved in the plant response to pathogen infection.

A principal object of the present invention is to provide additional means for chemically regulating the expression of a desired gene in a plant, seed, or plant tissue. In particular, the present invention relates to identification and isolation of a non-coding DNA sequence obtainable from the promoter region of an *Arabidopsis* PR-1a gene which, in the presence of chemical regulators, regulate the transcription of associated DNA sequences in plants.

Another principal object of the present invention is to provide transgenic plants, transgenic seeds and the progeny thereof expressing levels of plant pathogenesis-related anti-pathogenic proteins such as tobacco chitinase/lysozyme according to the invention, or substantially homologous proteins, which confer an enhanced disease-resistant and/or pest-resistant phenotype with respect to wild-type plants.

Accordingly, to meet this objective and others, the second aspect of the present invention disclosed herein provides for the isolation, cloning and identification of novel cDNA clones coding for plant pathogenesis-related (PR) proteins. These cDNAs, or their genomic counterparts, or DNA molecules with substantial homology to either (all of the above collectively referred to herein as "anti-pathogenic sequences"), can be engineered for expression of the encoded PR proteins or anti-sense RNA and transformed into plants to confer enhanced resistance or tolerance to various diseases and/or pests as described

herein. These DNA molecules may be engineered for constitutive expression, expression in particular tissues or at certain developmental stages, or induced expression in response to an inducer, for example in response to a chemical inducer as described herein.

The present invention further provides novel methods for differential screening and enriching for induced cDNAs, particularly those cDNAs induced in response to pathogen infection or a chemical inducer which triggers a response mimicking pathogen infection.

The present invention teaches the cloning of SAR genes by differential screening of tissues induced and non-induced to the systemic acquired response. SAR induction was found cause the transcription of genes in a protein synthesis-dependent fashion and also a protein synthesis-independent fashion. Two methods were used to clone specifically genes whose induced transcription is protein-synthesis independent:

Firstly, recombinant or chimaeric DNA molecules encoding genes which correspond to wild-type genes which can be chemically induced in a protein-synthesis independent manner, are obtained by

- (a) cloning genes which are chemically inducible by differential screening of cDNA libraries;
- (b) further analyzing the clones obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor; and
- (c) identifying clones which hybridise with both RNA chemically induced in the absence of a protein synthesis inhibitor and RNA chemically induced in the presence of a protein synthesis inhibitor.

Thus, cDNAs which were cloned by standard differential screening techniques were further screened on SAR-induced RNA isolated with and without cycloheximide (CHX) pre-treatment.

Secondly, recombinant or chimaeric DNA molecules encoding genes which correspond to wild-type genes which can be chemically induced in a protein-synthesis independent manner, are obtained by

- (a) cloning genes which are chemically inducible in the presence or absence of a protein synthesis inhibitor by differential display of isolated RNA using polymerase chain reaction;

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- (b) further analyzing the amplification products obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor; and
- (c) identifying amplification products which hybridise with both RNA chemically induced in the absence of a protein synthesis inhibitor and RNA chemically induced in the presence of a protein synthesis inhibitor.

Thus, a PCR-based "differential display" technique is used to identify SAR-induced, but protein synthesis independent cDNAs directly.

Differential display RNAs were prepared with and without SAR induction and CHX treatment. CHX as an inhibitor of protein synthesis is well known in the art and described by Greenberg *et al.*, *Mol. Cell Biol.* 6: 1050-1057 (1986), Lau and Nathans, *Proc. Natl. Acad. Sci.* 84: 1182-1186 (1987), and Uknes *et al.*, *Plant Cell* 5: 159-169 (1993). The methods described provide recombinant or chimaeric DNA molecules comprising plant SAR genes and preferably plant SAR cDNA molecules, wherein the wild-type genes corresponding to said SAR genes can be chemically induced in a plant in a protein-synthesis independent manner, that is in the presence of a protein synthesis inhibitor such as cycloheximide. Thus, a number of genes were cloned which were induced by the SAR response, yet expressed independently of protein synthesis. These cloned genes are likely signal transducers in the pathway leading from induction to the development of the resistant state.

Additionally recombinant or chimaeric DNA molecules are provided which comprise plant SAR genes and preferably plant SAR cDNA molecules, wherein the wild-type genes corresponding to said SAR genes can be chemically induced in a plant in a protein-synthesis dependent manner, that is only in the absence of a protein synthesis inhibitor such as cycloheximide. Recombinant or chimaeric DNA molecules encoding genes which correspond to wild-type genes which can be chemically induced in a protein-synthesis dependent manner, can be obtained by

- (a) cloning genes which are chemically inducible by differential screening of cDNA libraries;
- (b) further analyzing the clones obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor such as cycloheximide; and

- (c) identifying clones which hybridise with RNA chemically induced in the absence of a protein synthesis inhibitor but not with RNA chemically induced in the presence of a protein synthesis inhibitor.

Alternatively, these molecules can be obtained by

- (a) cloning genes which are chemically inducible in the absence of a protein synthesis inhibitor by differential display of isolated RNA using polymerase chain reaction;
- (b) further analyzing the amplification products obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor such as cycloheximide; and
- (c) identifying amplification products which hybridise with RNA chemically induced in the absence of a protein synthesis inhibitor but not with RNA chemically induced in the presence of a protein synthesis inhibitor.

DESCRIPTION OF GENE SEQUENCES

- SEQ ID No. 1: The cDNA sequence encoding an *Arabidopsis* class IV chitinase with a hevein domain (pChit4-TA).
- SEQ ID No. 2: The cDNA sequence encoding an *Arabidopsis* class IV chitinase without a hevein domain (pChit4-TB).
- SEQ ID No. 3: The cDNA sequence of the wheat gene WCI-1.
- SEQ ID No. 4: The partial cDNA sequence of the 5' end of the wheat gene WCI-2 which encodes a lipoxygenase isozyme. The partial sequence of the 3' end of this cDNA is provided in SEQ ID No. 5.
- SEQ ID No. 5: The partial cDNA sequence of the 3' end of the wheat gene WCI-2 which encodes a lipoxygenase isozyme. The partial sequence of the 5' end of this cDNA is provided in SEQ ID No. 4.
- SEQ ID No. 6: The cDNA sequence of the wheat gene WCI-3.
- SEQ ID No. 7: The cDNA sequence of the wheat gene WCI-4.
- SEQ ID No. 8: The cDNA sequence of the wheat gene WCI-5.
- SEQ ID No. 9: A tobacco protein-synthesis dependent gene involved in the regulation of the systemic acquired resistance response designated p1.1.1.

- SEQ ID No. 10: A tobacco protein-synthesis dependent gene involved in the regulation of the systemic acquired resistance response designated p11.31.4.
- SEQ ID No. 11: The 5' DNA sequence of a tobacco protein-synthesis independent gene involved in the regulation of the systemic acquired resistance response designated p11.30.13.
- SEQ ID No. 12: The DNA sequence of the 3' end of the same protein-synthesis independent gene described in Seq. ID. No. 11 cloned from tobacco and involved in the regulation of the systemic acquired resistance response designated p11.30.13. This sequence is derived from the non-coding strand (*i.e.* the "bottom" strand). The first base listed is therefore located in the furthest 3' position.
- SEQ ID No. 13: A tobacco protein-synthesis independent gene involved in the regulation of the systemic acquired resistance response designated p1.4.3. This sequence is identical to the thioredoxin gene published by Brugidou *et al.*, *Mol. Gen. Genet.* 238: 285-293 (1993).
- SEQ ID No. 14: A protein-synthesis independent SAR gene cloned from tobacco designated p66B1.
- SEQ ID No. 15: A protein-synthesis dependent SAR gene cloned from tobacco designated p14.22.3.
- SEQ ID No. 16: An *Arabidopsis* protein-synthesis independent gene involved in the regulation of the systemic acquired resistance response designated pDPA2.
- SEQ ID No. 17: The cDNA sequence encoding an *Arabidopsis* PR-1 protein cloned into plasmid pAPR1C-1.
- SEQ ID No. 18: Oligo B used in example 4A
- SEQ ID No. 19: Oligo C used in example 4A
- SEQ ID No. 20: The cDNA sequence encoding a basic tobacco chitinase/lysozyme protein cloned into the plasmid pBSCL2.
- SEQ ID No. 21: The cDNA sequence encoding an acidic tobacco chitinase/lysozyme protein cloned into the plasmid pBSTCL226.
- SEQ ID No. 22: A representative molecular adaptor sequence.
- SEQ ID No. 23: Oligonucleotide primer for the PR-1 gene.
- SEQ ID No. 24: Oligonucleotide primer for GUS gene.

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- SEQ ID No. 25: Oligonucleotide primer for the AHAS gene.
SEQ ID No. 26: Oligonucleotide primer for the BT gene.
SEQ ID No. 27: Peptide sequence used to design a degenerate forward oligonucleotide sequence for PCR cloning of chitinase IV
SEQ ID No. 28: Peptide sequence used to design a degenerate reverse oligonucleotide sequence for PCR cloning of chitinase IV
SEQ ID No. 29: The cDNA sequence of the Maize PR1-like gene PR-1mz
SEQ ID No. 30: Amino acid sequence of the protein encoded by SEQ ID No. 29
SEQ ID No. 31: The cDNA sequence of the Maize thaumatin-like gene PR-5mz
SEQ ID No. 32: Amino acid sequence of the protein encoded by SEQ ID No. 31
SEQ ID No. 33: The cDNA sequence of the Arabidopsis gene PSI-1
SEQ ID No. 34: The cDNA sequence of the Arabidopsis gene PSI-2
SEQ ID No. 35: The cDNA sequence of the Arabidopsis gene PSI-3
SEQ ID No. 36: The cDNA sequence of the Arabidopsis gene PSI-4
SEQ ID No. 37: The cDNA sequence of the Arabidopsis gene PSI-5

DEFINITIONS

In order to provide a clear and consistent understanding of the specification and the claims, including the scope given to such terms, the following definitions are provided:

Anti-pathogenic Sequence: A DNA molecule encoding a plant pathogenesis-related (PR) protein, or a DNA molecule with substantial homology thereto, which is capable of conferring enhanced resistance or tolerance to disease and/or pests when expressed in a plant, seed, or plant tissue.

Anti-sense Mechanism: A mechanism for regulation of gene expression based on the presence in a cell of a RNA molecule complementary to at least a portion of the mRNA encoded by the gene. This mechanism is thought to involve controlling the rate of translation of mRNA to protein due to the presence in a cell of an RNA molecule complementary to at least a portion of the mRNA being translated.

Associated DNA Sequence: A DNA sequence whose cellular activity either (1) regulates the activity of another DNA sequence or (2) is regulated by another DNA sequence. This definition specifically embraces, but is not limited to, sequences which are physically adjacent in a continuous DNA strand or which are physically separated. Physical separation includes, for example, separation within the same DNA strand, location within different DNA strands, or discontinuous interspersed sequences (e.g., alternating regulatable and coding sequences) in one strand.

Chemically Regulatable DNA Sequence: A DNA sequence which is capable of regulating the transcription of an associated DNA sequence where the regulation is dependent on a chemical regulator. The sequences may be of natural or synthetic origin.

Chemically Regulatable Gene: A gene containing at least one non-coding chemically regulatable DNA sequence and at least one associated coding DNA sequence. The genes may be of natural, synthetic or partially natural/partially synthetic origin.

Chemical Regulator (for a chemically regulatable DNA sequence): An elemental or molecular species which controls (e.g., initiates, terminates, increases or reduces), by direct or indirect action, the activity of a chemically regulatable DNA sequence in a system in which the chemical regulator is not normally found in an active form in an amount sufficient to effect regulation of transcription, to the degree and at the time desired, of a transcribable DNA sequence associated with the chemically regulatable DNA sequence. This terminology is intended to embrace situations in which no or very little regulator is present at the time transcription is desired or in which some regulator is present but increased or decreased regulation is required to effect more or less transcription as desired.

Thus, if the system containing the chemically regulatable DNA sequence is a plant, for example a transgenic plant, a chemical regulator is a species not naturally found in the plant in an amount sufficient to effect chemical regulation, and thus transcription of an associated gene, to the desired degree at the time desired.

By "direct action" it is intended that the chemical regulator action result from the direct interaction between the chemical regulator and the DNA sequence. By "indirect action" it is meant that the regulator action results from the direct interaction between the chemical regulator and some other endogenous or exogenous component in the system,

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the ultimate result of that direct interaction being activation or suppression of the activity of the DNA sequence. By "active form" it is intended that the chemical regulator be in a form required to effect control.

Chimeric Sequence or Gene: A DNA sequence containing at least two heterologous parts, e.g., parts derived from naturally occurring DNA sequences which are not associated in their naturally occurring states, or containing at least one part that is of synthetic origin and not found in nature.

Coding DNA Sequence: A DNA sequence which, when transcribed and translated, results in the formation of a cellular polypeptide.

Constitutive transcription: Transcription of substantially fixed amounts of a DNA sequence, irrespective of environmental conditions.

Gene: A discrete chromosomal region which is responsible for a discrete cellular product.

Inducers: Molecules that cause the production of larger amounts of macromolecules, compared to the amounts found in the absence of the inducer.

Inducible Protein: Proteins whose rate of production can be increased by the presence of inducers in the environment.

Non-coding DNA Sequence: A DNA sequence, which is not transcribed and translated, resulting in the formation of a cellular polypeptide when associated with a particular coding DNA sequence. A sequence that is non-coding when associated with one coding sequence may actually be coding when associated with another coding or non-coding sequence.

Phenotypic Trait: An observable property resulting from expression of a gene.

Plant Tissue: Any tissue of a plant in planta or in culture. This term includes, but is not limited to, whole plants, plant cells, plant organs, plant seeds, protoplasts, callus, cell cultures and any groups of plant cells organized into structural and/or functional units. The

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use of this term in conjunction with, or in the absence of, any specific type of plant tissue as listed above or otherwise embraced by this definition is not intended to be exclusive of any other type of plant tissue.

PR, or Pathogenesis-Related Proteins: Proteins expressed in plants reacting hypersensitively towards pathogens. This term embraces, but is not limited to, SAR8.2a and SAR8.2b proteins, the acidic and basic forms of tobacco PR-1a, PR-1b, PR-1c, PR-1', PR-2, PR-2', PR-2'', PR-N, PR-O, PR-O', PR-4, PR-P, PR-Q, PR-S, and PR-R major proteins, cucumber peroxidases, basic cucumber peroxidase, the chitinase which is a basic counterpart of PR-P or PR-Q, and the beta-1,3-glucanase (glucan endo-1,3- β -glucosidase, EC 3.2.1.39) which is a basic counterpart of PR-2, PR-N or PR-O, the pathogen-inducible chitinase from cucumber, class IV chitinases with and without a hevein domain, and the WCI ("Wheat Chemically Induced") gene proteins from . A hypersensitive reaction is characterized by a local necrosis of the tissues immediately surrounding the infection site of the pathogen and a subsequent localization of the pathogen, which is in contrast to a sensitive reaction wherein the pathogen spreads throughout the plant. Pathogens are, for example, viruses or viroids, e.g. tobacco or cucumber mosaic virus, ringspot virus or necrosis virus, pelargonium leaf curl virus, red clover mottle virus, tomato bushy stunt virus, and like viruses, fungi, e.g. Phytophthora parasitica or Peronospora tabacina, bacteria, e.g. Pseudomonas syringae or Pseudomonas tabaci, or aphids, e.g. Myzus persicae. This list is not limiting in any respect.

Regulation: The increasing (inducing) or decreasing (repressing) of the level of expression of a gene or the level of transcription of a DNA sequence. The definition is not intended to embrace any particular mechanism.

Substantially Pure DNA Sequence: A DNA molecule (sequence) isolated in substantially pure form from a natural or non-natural source. Such a molecule may occur in a natural system, for example, in bacteria, viruses or in plant or animal cells, or may be provided, for example, by synthetic means or as a cDNA. Substantially pure DNA sequences are typically isolated in the context of a cloning vector. Substantially pure means that DNA sequences other than the ones intended are present only in marginal amounts, for example less than 5%, less than 1%, or preferably less than 0.1%. Substantially pure DNA sequences and

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vectors containing may be, and typically are, provided in solution, for example in aqueous solution containing buffers or in the usual culture media.

Substantial Sequence Homology: Substantial sequence homology means close structural relationship between sequences of nucleotides or amino acids. For example, substantially homologous DNA sequences may be 80% homologous, preferably 90% or 95% homologous, and substantially homologous amino acid sequences may typically be 50% homologous, or more. Homology also includes a relationship wherein one or several subsequences of nucleotides or amino acids are missing, or subsequences with additional nucleotides or amino acids are interdispersed.

ABBREVIATIONS

The following abbreviations are used herein:

AHAS	acetoxyacid synthase
ATCC	American Type Culture Collection
ATP	adenosine triphosphate
bp	base pair
BT	<u>Bacillus thuringiensis</u> endotoxin
BTH	methyl benzo-1,2,3-thiadiazole-7-carboxylate
CAT	chloramphenicol acetyltransferase
CETAB	hexadecyltrimethylammonium bromide
CHX	cycloheximide
2,4-D	2,4-dichlorophenoxyacetic acid
DTT	dithiothreitol
dicamba	3,6-dichloro-2-methoxybenzoic acid
EDTA	ethylenediamine N,N,N',N'-tetraacetic acid
GUS	beta-1,3-glucuronidase
INA	isonicotinic acid
kb	kilo base pair
LUX	luciferase

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MES	2-(N-morpholino)ethanesulfonic acid
MU	4-methyl umbelliferyl glucuronide
NOS	nopaline synthase
NPT	neomycin phosphotransferase
NRRL	designation for deposits made with the Agricultural Research Culture Collection, International Depositing Authority, 1815 N. University Street, Peoria, Illinois 61604
OCS	octopine synthase
PEG	polyethylene glycol
picloram	4-amino-3,5,6-trichloropicolinic acid
PR protein	Pathogenesis-related protein
SA	salicylic acid
SAR	Systemic Acquired Resistance
SDS	sodium dodecyl sulfate
TFA	trifluoroacetic acid
TMV	tobacco mosaic virus
Tris-HCl	tris(hydroxymethyl)methylamine hydrochloride
WCI	Wheat Chemically Induced (gene nomenclature designation)

ANTI-PATHOGENIC SEQUENCES

The present invention also embraces anti-pathogenic DNA sequences which are capable of conferring enhanced disease resistance or disease tolerance when expressed in a plant or plant tissue. This includes coding sequences for plant pathogenesis-related (PR) proteins as described herein and sequences with substantial homology to these coding sequences.

Included within the scope of the present invention, in addition to the sequences exemplified specifically below and enumerated in the sequence listing, are cDNA sequences which are equivalent to the enumerated sequences which encode the given plant pathogenesis-related protein, and cDNA sequences which hybridize with the enumerated sequences and encode a polypeptide having some degree of disease-resistant activity of the given plant pathogenesis-related protein (i.e. an anti-pathogenic sequence).

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Equivalent cDNA sequences are those which encode the same protein even though they contain at least one different nucleotide from the enumerated sequence. As is well known in the art, the amino acid sequence of a protein is determined by the nucleotide sequence of the DNA. Because of the redundancy of the genetic code, i.e., more than one coding nucleotide triplet (codon) can be used for most of the amino acids used to make proteins, different nucleotide sequences can code for a particular amino acid. Thus, the genetic code can be depicted as follows:

Amino Acid	Codon	Amino Acid	Codon
Phenylalanine (Phe)	TTK	Histidine (His)	CAK
Leucine (Leu)	XTY	Glutamine (Gln)	CAJ
Isoleucine (Ile)	ATM	Asparagine (Asn)	AAK
Methionine (Met)	ATG	Lysine (Lys)	AAJ
Valine (Val)	GTL	Aspartic acid (Asp)	GAK
Serine (Ser)	QRS	Glutamic acid (Glu)	GAJ
Proline (Pro)	CCL	Cysteine (Cys)	TGK
Threonine (Thy)	ACL	Tryptophan (Trp)	TGG
Alanine (Ala)	GCL	Arginine (Arg)	WGJ
Tyrosine (Tyr)	TAK	Glycine (Gly)	GGL
Termination signal	TAJ		

Key: Each 3-letter deoxynucleotide triplet codon corresponds to a trinucleotide of mRNA, having a 5'-end on the left and a 3'-end on the right. All DNA sequences given herein are those of the strand whose sequence corresponds to the mRNA sequence, with thymine substituted for uracil. The letters stand for the purine or pyrimidine bases forming the deoxynucleotide sequence as follows:

A = adenine; G = guanine; C = cytosine; T = thymine

X = T or C if Y is A or G

X = C if Y is C or T

Y = A, G, C or T if X is C

Y = A or G if X is T

W = C or A if Z is A or G

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W = C if Z is C or T

Z = A, G, C or T if W is C

Z = A or G if W is A

QR = TC if S is A, G, C or T;

QR = AG if S is T or C

J = A or G

K = T or C

L = A, T, C or G

M = A, C or T

The above shows that the amino acid sequence of the instant plant pathogenesis-related proteins can be prepared using different nucleotide sequences encoding the same amino acid sequence of the proteins. Accordingly, the scope of the present invention includes such "equivalent nucleotide sequences."

cDNA sequences that hybridize with a given enumerated sequence and encode a polypeptide or protein having at least some degree of activity of the corresponding plant pathogenesis-related protein are those which exhibit substantial sequence homology, as defined hereinabove, with the enumerated Sequence such that it hybridizes with the latter under low stringency conditions. Such conditions are described in Examples 17 and 20 of European patent application EP-A-0 392 225. Proteins translated from these hybridizable cDNA sequences have different primary structures from proteins translated from the enumerated Sequences. However, their respective secondary structures are the same. Thus, the former proteins retain some degree of the biological activity (plant disease resistance) of the latter proteins and are considered as plant pathogenesis-related proteins also, e.g. "a PR-O protein," "a PR-N protein," etc. As used elsewhere herein, the term "activity" is referred to as the property of enhanced disease resistance or tolerance conferred upon a transgenic plant by virtue of its ability to express the DNA encoding the PR protein

It is contemplated by the invention that increased disease resistance or tolerance or increased scope of protection against a broader range of pathogens can be achieved by providing for the expression of more than one anti-pathogenic DNA sequence in a transgenic plant or plant tissue. The increased disease resistance thus achieved is due to a synergistic effect created by expressing a combination of anti-pathogenic sequences in the

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same plant or plant tissue. Thus, a method is provided which improves protection of a plant against a pest comprising transgenically expressing in said plant two or more DNA molecules encoding anti-pathogenic proteins, wherein the transgenically expressed proteins exert a synergistic effect. Preferably the DNA molecules encode anti-pathogenic proteins according to the invention. Due to synergy, the anti-pathogenic effect of a particular sequence may be more readily detected when combined with one or more other anti-pathogenic sequences than when present alone.

The present invention further includes chimeric genes and vectors comprising one or more anti-pathogenic sequences analogous to the chimeric genes and vectors comprising chemically regulatable sequences described in parts F and G above. Any desired promoter may be used in association with the anti-pathogenic sequences of the invention, including promoters which confer constitutive expression, promoters which confer expression in a particular tissue or subset of tissues and/or at a particular developmental stage, and promoters which confer chemically regulatable expression containing the noncoding sequences taught herein.

Also included as part of the present invention are plant tissues, plants and seeds comprising anti-pathogenic sequences analogous to those plant tissues, plants and seeds comprising chemically regulatable sequences described in part H above.

DIFFERENTIAL CLONING AND SCREENING TECHNOLOGY

A method has been conceived and developed which will allow efficient enrichment of sequences present in one population of molecules in greater amounts than in another population. The method's greatest utility is in situations where the populations are very similar and the differentially present sequences represent a very small proportion of the population.

If two populations of clones are similar and one wishes to isolate those clones which are present in one population in higher amounts (i.e. "induced" or differentially regulated"), past techniques involved screening with probes from the two populations (+/- screening; St. John and Davis, *Cell* 16:443-452 (1979)), or enrichment of probes or mRNAs by hybridization and hydroxy-apatite (HAP) chromatography (Davis, *et al.*, *Proc. Natl. Acad. Sci, USA* 81:2194-2198 (1984)). The first method has a demonstrated sensitivity limitation

in that only clones present in greater than about one in 2,000 will be detected. The second is laborious, technically difficult, and achieves enrichments of 20-50 fold at best.

The present method involves exploiting two recent developments in molecular technology: the polymerase chain reaction (Saiki *et al.*, *Science* 239:487-491 (1988)) and biotin-avidin chromatography [Stahl, *et al.*, *Nuc. Acids. Res.* 16: 3026-3038 (1988)]. The polymerase chain reaction (PCR) allows simple synthesis of large amounts of DNA of specified sequence. Biotin-avidin chromatography allows the efficient separation of molecules bearing a biotin affinity tag from those molecules which do not bear the tag.

In its general form, the technique consists of isolating single strands of cDNA representing two different populations ("induced" vs "uninduced"), but of opposite cDNA polarity for the two populations, i.e. one of "sense" polarity relative to mRNA's, and the other its complement, or "anti-sense", polarity relative to mRNA's. The isolated strands from the "induced" population would have no affinity tag, while the strands of opposite polarity from the "uninduced" populations would have stable affinity tags. When these two populations are hybridized together, hybrids will form between complementary strands present in the two populations. Those strands from the "induced" population which have no counterparts, or many fewer counterparts, in the "uninduced" population, remain single stranded.

Due to the presence of the affinity tag (in essence a handle) on the strands of the "uninduced" population molecules, those strands and, most importantly, any hybrid molecules can be removed from the mixture by affinity chromatography. This leaves only those "induced" molecules which are not significantly represented in the "uninduced" population. These "induced" molecules can then be cloned by standard means and serve as an enriched population from which to isolate "induced" clones; alternatively, the enriched molecules can be amplified individually and sequenced directly.

An alternate scheme is the same as described above except that it involves incorporating a labile affinity tag only on the "induced" population molecules, while the affinity tag on the "uninduced" population is stable. "Labile" in this case means that the affinity tag can be removed at will, or be altered at will in such a way that it no longer serves as an affinity tag. In this scheme all the molecules in the hybridization mixture could bind to the affinity matrix, but only those "induced" molecules that are not hybridized to a complementary "uninduced" counterpart could be selectively recovered from the matrix for subsequent cloning.

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The advantage of the methods of the invention described above over those previously described is the ability to isolate those genes which are turned on only to low levels, in specific circumstances, and which may play a causative role in some important biological phenomenon.

The present invention teaches the cloning of SAR genes by differential screening of tissues induced and non-induced to the systemic acquired response SAR induction causes the transcription of genes in a protein synthesis-dependent fashion and also a protein synthesis-independent fashion. Two methods were used to clone specifically genes whose induced transcription is protein-synthesis independent. Firstly, cDNAs which were cloned by standard differential screening techniques were further screened on SAR-induced RNA isolated with and without cycloheximide (CHX) pre-treatment. Secondly, a PCR-based "differential display" technique was used to identify SAR-induced, but protein synthesis independent cDNAs directly. Differential display RNAs were prepared with and without SAR induction and CHX treatment. The use of CHX as an inhibitor of protein synthesis is well known in the art and is described by Greenberg *et al.*, *Mol. Cell. Biol.* 6: 1050-1057 (1986), Lau and Nathans, *Proc. Natl. Acad. Sci.* 84: 1182-1186 (1987), and Uknes *et al.*, *Plant Cell* 5: 159-169 (1993). Thus, a number of genes were cloned which were induced by the SAR response, yet expressed independently of protein synthesis. These cloned genes are likely signal transducers in the pathway leading from induction to the development of the resistant state.

EXAMPLES**Example (i): *Ligation In Agarose***

Following restriction digestion of plasmid DNA and electrophoretic separation of the fragments on a low melting TAE gel, the bands containing appropriate fragments are precisely excised and heated to 65°C to melt the agarose 2-5 µl are added to 15 µl water and the solution is left at 65°C for 10 minutes. This solution is cooled to 37°C and left for five minutes to equilibrate to temperature 2 µl of 10 X ligase buffer (200 mM Tris, pH 8.0, 100 mM MgCl₂, 100 mM DTT, 10 mM ATP) are added along with 1 µl T4 DNA ligase (New England BioLabs), and this solution is allowed to solidify and incubate at 15°C overnight.

Example (ii): *Transformation From Agarose*

The agarose containing the appropriate DNA is melted by incubating at 65°C for 10 minutes 10 µl of this solution are added to 30 µl of TE buffer (10 mM Tris pH 7.5, 1 mM EDTA), mixed and allowed to stand at room temperature. Frozen competent cells (*E. coli* strain DH5) are placed on wet ice to thaw. The diluted DNA solution is added to 200 µl of cells and allowed to stand on ice for 20 minutes. The cells containing the DNA are then heat-shocked for 90 seconds at 41°C. The cells are then left at room temperature for 10 minutes 0.8 ml of SOC medium (Hanahan, D., *J. Mol. Biol.* 166: 557-580 (1983)) is added and the culture is incubated at 37°C for one hour 100 µl of the culture is plated on LB plates (Miller, *supra*) containing 100 µg/ml ampicillin (L-amp) and the plates are incubated overnight at 37°C. Positive colonies are picked and restreaked to a second L-amp plate and the plates are incubated overnight at 37°C.

Example (iii): *Labelling DNA Restriction Fragments*

DNA is treated with the appropriate restriction enzymes and fragments are separated by electrophoresis on a low-gelling temperature agarose gel. A band containing the fragment of interest is excised and the DNA purified by standard techniques 50 ng of the DNA fragment is labelled using the IBI Random primer kit "Prime time" according to the manufacturers directions.

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Example (iv): Southern Blotting

3 µg of tobacco DNA is digested with various restriction enzymes under the conditions suggested by the supplier. The DNA is extracted with phenol, precipitated with ethanol and then resuspended in gel loading buffer (15% ficoll, 0.025% bromophenol blue, 10 mM EDTA, pH 8). Samples are loaded and electrophoresed on a 0.5% agarose gel at 5 V/cm until the bromophenol blue dye reaches the end of the gel. The DNA is transferred to Gene-Screen Plus (DuPont) using the alkaline transfer procedure as described by the supplier. Pre-hybridization, hybridization and washing are according to the manufacturer's recommendation. Hybridization is detected by autoradiography.

Example (v): Molecular Adaptors

A typical molecular adaptor for the conversion of a PstI site to a BamHI site is the sequence

5'-GGGATCCCTGCA-3' (SEQ ID No. 22).

This molecule is synthesized on an Applied Biosystems Synthesizer using B-cyano-ethylphosphoramidite chemistry and purified by reverse-phase HPLC. About 2 µg of this oligonucleotide is kinased according to Maniatis *et al.*, supra, p. 125. The oligonucleotide solution is heated to 65°C in a water bath and allowed to cool to room temperature over a period of about 30 minutes. An approximately 10-fold molar excess of this annealed adapter is added to the digested DNA along with 10 X ligase buffer, T4 DNA ligase, and an appropriate amount of water. A typical reaction is:

DNA to be adapted:	1-2 µl (~ 1 pmol)
Adapter:	1 µl (~10 pmol)
10 X ligase buffer:	1 µl
T4 DNA ligase:	1 µl
Water:	5-6 µl

This solution is incubated at 12-15°C for 30 minutes, and heated to 65°C for 30 minutes to inactivate the ligase. The salt concentration and volume are adjusted for the appropriate restriction digest and the adapted DNA is digested to expose the adapted

"sticky end." Unincorporated adapters are removed either by electrophoresis on an agarose gel or by sequential isopropanol precipitations.

Example (vi): *Primer Extension Mapping*

A. Synthesis and 5' End Labeling of Primers for Primer Extension

The following primer oligomers are synthesized using an Applied Biosystems Synthesizer and β -cyanoethylphosphoramidite chemistry:

PR-1: 5'-ATAGTCTTGTTGAGAGTT-3' (SEQ ID No. 23)

GUS: 5'-TCACGGGTTGGGGTTTCTAC-3' (SEQ ID No. 24)

AHAS: 5'-AGGAGATGGTTTGGTGGA-3' (SEQ ID No. 25)

BT: 5'-ATACGTTCTACTATCATAGT-3' (SEQ ID No. 26)

The oligonucleotides are purified by reverse-phase high pressure liquid chromatography (HPLC) 5 pmol of each oligo is kinased (Maniatis, T. *et al.*, supra, at p. 125) using 200 μ C of 32 P-ATP (6000 Ci/mmol, 10 μ Ci/ μ l). After incubation at 37°C for 30 minutes, the reaction is diluted to 100 μ l, extracted with phenol/chloroform and then precipitated three times with 50 μ g carrier RNA. The final precipitate is resuspended in 1 X reverse-transcriptase buffer (50 mM Tris-HCl, pH 7.5, 40 mM KCl, 3 mM $MgCl_2$) at a concentration of 2 nM. The specific activity of the labeled oligonucleotide is determined to be about 3×10^6 Cvcpm/pmol.

B. Total RNA Preparation

Total RNA is prepared essentially as described by Lagrimini, L.M. *et al.*, *Proc. Natl. Acad. Sci. USA* 84: 7542 (1987). Tissue is ground under liquid nitrogen in a mortar and pestle. The ground tissue is added to grinding buffer (Lagrimini *et al.*, supra) using 2.5 ml per gram tissue. An equal volume of phenol is then added and the emulsion is homogenized in a Brinkman polytron. A one-half volume of chloroform is added and the emulsion is gently mixed for 15 minutes. The phases are separated by centrifugation and

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the aqueous phase is removed. RNA is precipitated by the addition of sodium acetate to 0.3 M and 2.5 volumes ethanol. The precipitate is collected by centrifugation and resuspended in 2 ml sterile water. Lithium chloride is added to a final concentration of 3 M and left at 4°C overnight. The precipitate is collected by centrifugation and the pellet is washed with ice-cold 80% ethanol. The pellet is dried and resuspended in 500 µl sterile water. The concentration of this total RNA preparation is determined spectrophotometrically.

Alternatively, RNA is extracted from callus as described above except that the callus tissue is cut into cubes approximately 3 mm in size, and added to pre-chilled mortars and pestles for grinding in liquid nitrogen prior to the polytron step.

C. Primer Extension

50 µg of total RNA is lyophilized in a 500 µl Eppendorf tube. The RNA is resuspended in 30 µl of radiolabeled probe solution and heated to 70°C for 10 minutes. The tube is slowly cooled to 37°C and allowed to incubate overnight. Without removing the tube from the 37°C water bath, 2 µl of 10 X reverse-transcriptase buffer (500 mM Tris-HCl, pH 7.5, 400 mM KCl, 30 mM MgCl₂), 1 µl 5 mg/ml bovine serum albumin, 5 µl 100 mM dithiothreitol, 5 µl 10 X dNTPs (10 mM of each dNTP in H₂O), 3 µl H₂O, 2 µl RNasin (80 units), and 2 µl reverse transcriptase (400 units) are added and the reaction is incubated at 37°C for 30 minutes. To stop the reaction, 5 µl of 3 M sodium acetate, pH 5, and 150 µl absolute ethanol are added. The tube is left at -20°C for 30 minutes, the precipitate is collected by centrifugation, washed with 80% ethanol and allowed to air-dry. The precipitate is resuspended in 10-20 µl of loading dye (90% formamide, 0.05% bromophenol blue, 0.05% xylene cyanol, 1 mM EDTA) and the extension products are separated on a 6% sequencing gel (Maniatis, T. *et al.*, supra). Extension products are visualized by autoradiography.

Example (vii): S1 Nuclease Mapping

The plasmid pBS-PR1013Cla is digested with SfaNI, dephosphorylated with calf intestinal phosphatase and kinased with ³²P-ATP. Following phenol extraction and ethanol precipitation, the DNA is digested with BstEII and the 300 bp fragment from 750 to 1035 of Figure 1 is isolated after electrophoresis on a low gelling temperature agarose gel. The probe is resuspended in formamide hybridization buffer (Berk, A.J. *et al.*, Cell 12, 721

(1977)) at a concentration of about 2 nM. The specific activity of the probe is about 5×10^6 Cpm/pmol.

Lyophilized, total RNA (50 μ g) is dissolved in 30 μ l of the probe solution, and the tubes are heated to 65°C for 10 minutes, then allowed to hybridize overnight at 48°C. S1 nuclease treatment and gel electrophoresis are essentially as described, using an S1 concentration of 400 units/ml and an incubation temperature of 30°C. The appropriate S1 nuclease concentration is determined in pilot experiments.

Example (viii): Mapping the Transcriptional Start Site

The transcriptional start site for the PR-1a gene is determined by a combination of S1 nuclease mapping and primer extension analysis. An autoradiogram of a primer extension experiment using either RNA isolated from TMV-infected leaves or an mpl9 subclone of the XhoI-PstI fragment as a template and a 17 base oligonucleotide complementary to positions 1025 to 1042 of the PR-1a sequence as a specific primer is examined. The primer itself is labeled at the 5' phosphate, therefore the size of the extension product will be identical to the size of the corresponding band in the sequencing lanes. The appearance of two strong bands corresponding to positions 902 and 903 and a weak band at position 901 of the genomic clone suggests transcription initiating at either of these positions. However, primer extension analysis alone cannot be used to identify the 5' end of a mRNA. For instance, the mRNA may contain a 5' end that has been spliced from an upstream location.

To determine conclusively the 5' end, high resolution S1 nuclease mapping is used in conjunction with primer extension. An SfaNI fragment is labeled at the 5' end and digested with BstEII to yield a strand specific probe extending from position 759 to 1040. This probe is used to map the 5' end of PR-1a transcripts in RNA isolated from TMV-infected tobacco leaves. A major band of 137-2 bases is found which corresponds to positions 901 to 905 of the genomic clone. In high resolution S1 experiments, where the digestion products are electrophoresed along with a size standard of sequencing reactions performed on the probe, three bands are visualized corresponding to positions 901, 902 and 903. These results confirm the primer extension analysis and place the 5' end of the PR-1 mRNA at either position 901, 902 or 903. With regard to transcription initiation, one possible interpretation of these results is that RNA polymerase begins transcription at either base 901, 902 or 903 with more or less equal probability. However, since eukaryotic

transcription favors initiation at an A, a more likely explanation for the apparent multiple 5' ends is that the PR-1a mRNA begins at position 903 (an A) and the PR-1b and -1c mRNAs begin each at one of the other positions on their corresponding genes.

Example (ix): General Techniques for Peptide Generation, Purification, and Automated Sequencing

The proteins relevant to these examples are isolated, purified and sequenced in some cases, for the first time and in accordance with literature procedures in other, for the purpose of allowing the isolation of the corresponding cDNA's and ultimately for confirming the identities of their cDNA's and chemically inducible genes.

A. Reduction and Alkylation

Purified, lyophilized protein is dissolved in 6 M guanidine-HCl containing 1 M Tris-HCl, pH 8.6, 10 mM EDTA. Dithiothreitol is added to 20 mM and 4-vinylpyridine is added to a final concentration of 50 mM. The sample is then incubated for 1.5 hours under nitrogen. The pyridylethylated material is desalted on HPLC using an Aquapore phenyl column (2.1 x 10 cm, Brownlee). The column is eluted with a linear, 5-80% gradient of acetonitrile/isopropanol (1:1) in 0.1% trifluoroacetic acid (TFA).

B. Cyanogen Bromide Cleavage and Removal of Pyroglutamate

Cyanogen bromide cleavage is performed in situ according to Simpson, R.J. *et al.*, *Biochem. Intl.* 8: 787 (1984) Digestion of PR-1 protein with pyroglutamate aminopeptidase (Boehringer Mannheim) is carried out according to Allen, G., *Plant Sci. Lett.* 26: 173 (1982).

C. LysC digestion

Protein is digested with endoproteinase Lys-C (Boehringer Mannheim) in 0.1 M Tris-HCl, pH 8.5, for 24 hours at room temperature using an enzyme:substrate ratio of 1:10. Resulting peptides are isolated by HPLC using an Aquapore C-8 column (1 x 22 cm, Brownlee) eluted with a linear acetonitrile/isopropanol (1:1 ratio) gradient (0 to 60%) in 0.1% TFA.

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D. Trypsin Digestion

Digestion with trypsin (Cooper) is performed in 0.1 M ammonium bicarbonate, pH 8.2, containing 0.1 M calcium chloride for five hours at 37°C using an enzyme:substrate ratio of 1:100. Peptides generated are separated on HPLC using the same conditions as with the Lys-C peptides or performed in 0.1 M Tris-HCl pH 8.5 for 24 hours at 37°C using an enzyme to substrate ratio of 1:50. Peptides are isolated by HPLC using a Vydac C-18 column (2.1 x 150 mm) with a linear 0 to 60% acetonitrile:isopropanol (1:1) gradient in 0.1 % TFA.

E. Sequencing

Automated Edman degradations are performed with an Applied Biosystems 470A gas-phase sequencer. Phenylthiohydantoin (PTH) amino acids are identified using an Applied Biosystems 120A PTH analyzer.

Example 1A: Cloning of cDNAs corresponding to SAR CHX-independent genes from tobacco

A number of cDNAs were cloned by differential screening from cDNA prepared from induced and non-induced tissue. The induced cDNA was prepared from tobacco leaves which had been pre-treated with methyl benzo-1,2,3-thiadiazole-7-carboxylate (BTH), whereas the non-induced cDNA was prepared from tissue which had not been pre-treated with BTH. cDNA libraries were prepared in λ ZAP II (STRATAGENE). A standard differential screening technique was used. Plaques carrying induced cDNA were plated at low density and transferred to two sets of hybridization filters. Known SAR gene sequences were hybridized to the first filter and uninduced cDNA to the second. The second filter was then stripped and hybridised with induced cDNA. Plaques which hybridized with the induced cDNA probe, but not with the uninduced cDNA probe or with known SAR gene sequences were potential novel SAR genes and were picked directly for further analysis. Plating had been at a plaque density which was sufficiently low to enable these plaques to be picked as nearly pure plaques. Multiple candidates for each plaque were *in vivo* excised, according to the manufacturer's recommended conditions, for further screening by Northern hybridization to RNA isolated from either untreated or BTH-treated tobacco plants. Individual clones chosen from the secondary screen were further analyzed by Northern

hybridisation to RNA isolated from tobacco plants which had been pre-treated with salicylic acid (3 mM), INA (1 mM), and BTH (1 mg/ml), all in the presence or absence of cycloheximide (CHX; 1 mg/ml). Inducer pre-treatments were done at 2 h, one day, and eight days before the isolation of RNA, whereas CHX treatment was done at one day before isolation of RNA. Three cDNAs were found induced in a protein synthesis independent fashion. The genes corresponding to these cDNAs have been designated p66B1 (SEQ ID NO. 14), p11.30.13 (SEQ ID NO. 11), and p1.4.3 (SEQ ID NO. 12 / SEQ ID NO. 13) and are likely signal transducers of the SAR response. Gene p1.4.3 has previously been disclosed as a thioredoxin (Brugidou *et al.*, *Mol. Gen. Genet.* 238: 285-293 (1993)). However, this is the first disclosure of the gene's likely involvement in the systemic acquired response. Three cDNAs were found to be expressed in a protein synthesis dependent fashion. These were designated p1.1.1, p11.31.4, and p14.22.3 and are listed as SEQ. ID Nos. 9, 10, and 15, respectively.

Example 1B: Cloning of cDNAs corresponding to SAR CHX-independent genes from Arabidopsis using INA for induction

Total RNA was isolated from the following Arabidopsis lines: (1) untreated, (2) INA treated (0.25 mg/ml), (3) CHX treated (1 mg/ml), and (4) INA + CHX treated. Treatments were made 1 day before RNA isolation. RNA thus isolated was subjected to "differential display" using the protocol described by Liang and Pardee, *Science* 257: 967-971 (1992). Amplified fragments which were found in both the INA as well as the INA + CHX treated RNA samples were gel-purified and used as probes on Northern blots carrying similarly induced RNA samples. Fragments for which Northern hybridization confirmed the induction profile apparent from differential display were subcloned into a plasmid vector. Using the cloned fragment it was possible to isolate near full-length cDNAs from a cDNA library produced by BTH induction:

Mature Arabidopsis thaliana ecotype Columbia (Lehle Seeds, Tucson, AZ) plants are sprayed with a 0.5 mg/ml suspension of a wettable powder formulation of methylbenzo-1,2,3-thiadiazole-7-carboxylate consisting of 25% active ingredient. Seven days later, the leaf tissue is harvested and frozen in liquid N₂. Total RNA is isolated as described in Example (vi) and Poly (A)⁺ RNA is isolated using a Poly (A) QuikTM mRNA isolation kit from Stratagene (La Jolla, CA). This Poly (A)⁺ RNA is then used to make a cDNA library in the

uni-zapTM XR vector (Stratagene) using a ZAP-cDNATM Gigapack^R II Gold cloning kit from Stratagene. A portion of the cDNA library is amplified as described in the Stratagene kit.

The cDNA pDPA2 was cloned using this technique and is induced by the SAR response in a protein synthesis independent fashion. Its sequence is listed in Seq. ID No. 16.

Example 1C: Cloning of cDNAs corresponding to SAR CHX-independent genes from Arabidopsis using BTH for induction

RNA isolated from (1) untreated, (2) BTH treated, and (3) BTH + CHX treated Arabidopsis lines was subjected to "differential display" using the protocol described by Liang and Pardee, *Science* 257: 967-971 (1992) and as described in example 1B above. Amplified fragments which were found in both the BTH as well as the BTH + CHX treated RNA samples were gel-purified and used as probes on Northern blots carrying similarly induced RNA samples. Fragments for which Northern hybridization confirmed the induction profile apparent from differential display were subcloned into a plasmid vector. The following clones induced in a protein synthesis-independent manner were obtained:

- PSI-1 (SEQ ID NO. 33): cDNA homologous to EMBL Accession #Z26429, an *A. thaliana* transcribed sequence (clone FAFC80-1; 5' end and actin depolymerizing factor from *L. longiflorum*), and EMBL Accession #Z14110, an *L. longiflorum* mRNA for actin depolymerizing factor
- PSI-2 (SEQ ID NO. 34): cDNA with no known homology
- PSI-3 (SEQ ID NO. 35): cDNA homologous to EMBL Accession # T20772, *A. thaliana* cDNA 92H8T7, and EMBL Accession #T04101, *A. thaliana* cDNA clone SCG7T7P
- PSI-4 (SEQ ID NO. 36): cDNA homologous to EMBL Accession #D22118, a partial Rice cDNA sequence
- PSI-5 (SEQ ID NO. 37): cDNA with no known homology

Example 2A: Cloning of Class IV Chitinase cDNAs

To clone recombinant or chimaeric DNA molecules encoding class IV chitinase class specific degenerate oligonucleotides were designed from areas of homology between bean PR4 chitinase (Margis-Pinheiro *et al.*, *Plant Mol. Biol.* 17: 243-253 (1991)) and sugar beet chitinase 4 (Mikkelsen *et al.*, in "Advances in Chitin and Chitosan, ed. by Brine *et al.*, pub. by Elsevier, Amsterdam (1992)): oligonucleotides were designed degenerate for the peptide

sequences HFCYIEE (forward, spanning nucleotides 406-426; SEQ ID NO. 27), and IRAING (reverse, spanning nucleotides 705 to 675; SEQ ID NO. 28). DNA was extracted from two-week old plants of *Arabidopsis thaliana* ecotype Landsberg and amplified using a Perkin-Elmer thermal cycler 480 at the following cycle settings: 94°C for 5 minutes; 35 cycles at 94°C, 1 minute, 43°C or 45°C, 1 minute, and 72°C, 2 minutes; followed by 5 minutes at 72°C. The amplified fragment was gel purified, collected by centrifugation through Whatman paper, ethanol precipitated, resuspended in TE, digested with BamHI and NsiI and subcloned into pTZ18U (Pharmacia). Four clones of the fragment were sequenced; they differed only within the oligo-derived sequence as could be expected from amplification with degenerate oligonucleotides.

Two different cDNAs were isolated simultaneously by screening a leaf tissue cDNA library (Uknes *et al.*, *Plant Cell* 4: 645-656 (1992)) at high stringency with the PCR amplified genomic fragment described above. Duplicate plaque lifts were taken with nitrocellulose filters (Schleicher & Schuell, Keene, NH) (Ausubel *et al.*, in "Current Protocols in Molecular Biology, pub. by J. Wiley & Sons, New York (1987)). Probes were labelled by random priming (using the labeling system supplied by Gibco BRL, Gaithersburg, MD). Hybridization and washing were done at 65°C according to Church and Gilbert, *Proc. Natl. Acad. Sci.* 81: 1991-1995 (1984). Positive plaques were purified and plasmids containing the cDNA inserts were *in vivo* excised for DNA sequence determination. Of the six positive clones two contained an insert with structural homology to previously characterized class IV chitinases and were designated class IV chitinase type A and four clones were divergent in that they lacked the class IV chitinase hevein domain; these were designated class IV chitinase type B. As none of the cDNA inserts was full-length, an additional 29 bp of class IV chitinase type A and 17 bp of class IV chitinase type B, both containing a methionine initiation codon were amplified from ethephon-induced RNA using the 5' RACE system for rapid amplification of cDNA ends (Gibco BRL, Gaithersburg, MD). Sequence comparisons were performed using the GAP and PILEUP features of the Genetics Computer Group software (Genetics Computer Group, Madison, WI).

Class IV chitinase type A is a 1079 base pair cDNA with an open reading frame of 264 amino acids containing the characteristic cysteine-rich hevein and chitinolytic domains and the three short deletions typically found in class IV chitinases (see SEQ ID No. 1). The cDNA for class IV chitinase type B is 952 base pairs in length and encodes a protein of 214 amino acids which lacks a hevein domain and contains a fourth deletion (see SEQ ID No.

2). The cDNAs are 71% identical overall and 80% identical over coding sequence. The predicted protein encoded by Arabidopsis class IV chitinase type A is 89% homologous to the basic Brassica napus (rapeseed) class IV chitinase, 61% homologous to basic Beta vulgaris (sugar beet) class IV chitinase, 57% homologous to basic Zea mays class IV chitinase B, 58% homologous to acidic Phaseolis vulgaris (bean) PR4 class IV chitinase, and 55% homologous to acidic Dioscorea japonica (yam) class IV chitinase. It is 42% homologous to Arabidopsis thaliana basic class I chitinase.

The predicted mature protein encoded by Arabidopsis class IV chitinase type A has a molecular weight of 25695 D and a pI of 7.8; whereas the protein encoded by Arabidopsis class IV chitinase type B has a molecular weight of 20553 D and a pI of 10 assuming the removal of a signal peptide based on homology to tobacco PR-Q.

Northern analysis showed that both chitinases were induced by TCV infection, confirming their classification as PR-proteins.

Using techniques well known in the art, these cDNAs can be cloned into expression cassettes and vectors for transfer to transgenic plants. Typical techniques used in the art are described in section 4A (examples 22 to 35), section 5A (examples 36 to 56) and section 6 (examples 57 to 80) of EP-A 0 392 225.

Example 2B: Isolation of Chemically Induced Wheat cDNAs Using the Method of Differential Plaque Filter Hybridization

Recombinant or chimaeric DNA molecules comprising the cDNA of a wild-type wheat gene which wild-type gene can be chemically induced in a plant were obtained in the following way:

Samples of winter wheat (cultivar Kanzler) were harvested 2-3 days after treatment with either water or 200 ppm of the plant activator compound benzo-1,2,3-thiodiazole-carboxylic acid. Total RNA was prepared from frozen tissue samples using a standard phenol extraction/LiCl precipitation procedure (Lagrimini *et al.*, *Proc. Natl. Acad. Sci.* 84: 7542-7546 (1987)). PolyA (+) RNA was purified from total RNA using the Poly(A) Quik mRNA purification kit (Stratagene Cloning Systems, LaJolla, CA). A bacteriophage lambda ZAP II cDNA library was prepared from the benzo-1,2,3-thio-diazolecarboxylic acid treated polyA(+) sample using the Uni-Zap XR Gigapack II Gold cloning kit (Stratagene) as described by the manufacturer. The phage library was plated at a density of approximately 5000 plaques on a 10 cm petri dish and grown for 6-8 hours at 37°C. Duplicate filter lifts of

the plaques were made using nitrocellulose membranes (Schleicher & Schuell, Keene, NH). Labelled first strand cDNA probes were prepared from polyA of both the water control and the benzo-1,2,3-thiodiazolecarboxylic acid-treated samples using ^{32}P -dCTP and the AMV reverse transcriptase (GibcoBRL, Gaithersburg, MD) under the manufacturer's conditions. Each probe was hybridized ($>10^6$ cpm/ml) with one set of the duplicate lifts overnight at 65°C . Hybridization and wash conditions were as described in Church and Gilbert, *Proc. Natl. Acad. Sci.* 81: 1991-1995 (1984). Hybridization was detected by autoradiography.

Plaques appearing to hybridize preferentially to the chemically treated cDNA were purified and their cDNA inserts were amplified using the GeneAmp Polymerase Chain Reaction (PCR) kit (Perkin Elmer, Norwalk, CT) and primers homologous to the flanking lambda Zap II sequences. The amplified inserts were excised from a low melting temperature SeaPlaque GTG agarose gel (FMC BioProducts, Rockland, ME) and labelled using ^{32}P -dCTP and the Random Primers DNA Labeling System (Gibco BRL). These probes were hybridized with total RNA blots (Ausubel *et al.*, in "Current Protocols in Molecular Biology; pub. by J. Wiley & Sons, New York (1987)) of control and benzo-1,2,3-thiodiazolecarboxylic acid treated RNAs to verify that they contained chemically induced cDNAs.

The induced clones were *in vivo* excised into pBluescript plasmids according to the manufacturer's instructions (Stratagene) and plasmid DNAs were purified using Magic Miniprep columns (Promega Biotech, Madison, WI). The cDNA sequences were determined by the chain termination method using dideoxy terminators labelled with fluorescent dyes (Applied Biosystems, Inc., Foster City, CA). The DNA and the predicted amino acid sequences were compared to available databases using the GAP (Deveraux *et al.*, *Nucl. Acids Res.* 12: 387-395 (1984)) and the BLAST (Altschul *et al.*, *J. Mol. Biol.* 215: 403-410 (1990)) programs.

The nucleotide sequence of one apparently full length induced clone is set forth in SEQ ID No. 3. This clone, denoted WCI-1 (Wheat Chemical Induction), was found to share limited homology with two rice cDNAs, one expressed specifically in the shoot apical meristem (De Pater and Schilperoort, *Plant Mol. Biol.* 18: 161-164 (1992)), the other inducible by salt stress (Claes *et al.*, *Plant Cell* 2: 19-27 (1990)). The function of these proteins is unknown.

The predicted amino acid sequence of a second highly induced cDNA, WCI-2, clearly identified it as an isozyme of wheat lipoxygenase, based on its homology to other

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plant lipoxygenases in the database. The DNA sequence of this cDNA is shown in SEQ ID No. 4.

A third class of induced cDNA (WCI-3) was isolated which to date shows no significant homology to sequences in the databases. The DNA sequence of this apparently full-length clone is set forth in SEQ ID No. 6.

Using techniques well known in the art, these cDNAs can be cloned into expression cassettes and vectors for transfer to transgenic plants. Typical techniques used in the art are described in section 4A (examples 22 to 35), section 5A (examples 36 to 56) and section 6 (examples 57 to 80) of EP-A 0 392 225.

Example 2C: Isolation of Wheat cDNAs Specifically Induced by Treatment With benzo-1,2,3-thiodiazolecarboxylic acid, Using the Method of Differential cDNA Display

The total and polyA(+) samples described in Example (vi) were used for PCR differential display of the mRNA from water and benzo-1,2,3-thiodiazolecarboxylic acid treated wheat tissue essentially as described in Liang and Pardee, *Science* 257: 967-971 (1992). Amplified cDNA fragments that appeared to be present only in the chemically treated sample were excised from the dried sequencing gel and electroeluted using a Centrilotur device and Centricon-30 Microconcentrators (Amicon, Beverly, MA). The purified fragments were PCR amplified using primers that consisted of the original differential display 10-mers plus an additional 10 bases of unique sequence added to their 5' ends. After 8 PCR cycles at low annealing temperature (42-45°C), the annealing temperature was raised to 60°C for an additional 30 cycles. Essentially 100% of the fragments could then be visualized by EtBr staining and were excised from a SeaPlaque GTG agarose gel (FMC).

These gel fragments were labelled and used to probe RNA blots as previously described. Fragments that hybridized only with chemically treated RNA were TA-cloned into the plasmid vector pCR II using a TA Cloning Kit (Invitrogen Corporation, San Diego, CA). Inserts from the plasmids were screened again against control/chemical RNA blots to verify that the inducible gene fragment had been subcloned. Induced fragments were then labelled by random priming and hybridized against filter lifts of the chemically induced cDNA library as described in Example 2A. Hybridizing plaques were purified, sequenced, and analyzed as previously described in order to obtain full length clones corresponding to the

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original small (200-400 bp) fragments, and to identify the induced gene product where possible.

The nucleotide sequence of one clone obtained by this procedure is set forth in SEQ ID No. 7. This clone, WCI-4, has some homology to known thiol protease sequences from a variety of sources and may therefore be a thiol protease.

Partial sequences of an additional induced gene that was isolated using differential display is set forth in SEQ ID No. 8. This fragment shows no database homology and has been designated WCI-5.

Using techniques well known in the art, these cDNAs can be cloned into expression cassettes and vectors for transfer to transgenic plants. Typical techniques used in the art are described in section 4A (examples 22 to 35), section 5A (examples 36 to 56) and section 6 (examples 57 to 80) of EP-A 0 392 225.

Example 3A: Preparation of cDNA Library From TMV-Infected Tobacco Leaves.

Nicotiana tabacum cv. Xanthi-nc leaves are infected with tobacco mosaic virus and harvested five days post-infection. Total RNA is prepared as described above and poly A+ RNA is isolated using standard techniques. A cDNA library is constructed using this poly A+ RNA in the Lambda onC cloning vector (Stratagene) essentially as described (Gubler, U. and Hoffman, B. J., *Gene* 25:263 (1983)).

Example 3B: Isolation of cDNA Clones Encoding Chitinase/Lysozymes

To clone recombinant or chimaeric DNA molecules encoding chitinase/lysozyme about 300,000 plaques of the TMV-infected tobacco cDNA library described in Example 3A are screened using a labeled cDNA probe encoding the cucumber chitinase/lysozyme cDNA and washing filters at 50°C in 0.125 mM NaCl, 1% SDS, 40 mM sodium phosphate (pH 7.2), 1 mM EDTA. Positive plaques are purified and the DNA sequence of two clones, named pBSCL2 and pBSTCL226 are determined. These are presented in SEQ ID Nos. 20 and 21, respectively. The proteins encoded in the clones of these sequences are determined to be chitinase/lysozymes based on structural homology to the cucumber chitinase/lysozyme.

In addition, a protein is purified from intercellular fluid of TMV-infected tobacco. Peptides are generated and sequenced as described in Example (ix)

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The protein encoded by pBSTCL226, corresponding to an acidic isoform of chitinase/lysozyme, was found to match the deduced peptide sequences exactly.

Using techniques well known in the art, these cDNAs can be cloned into expression cassettes and vectors for transfer to transgenic plants. Typical techniques used in the art are described in section 4A (examples 22 to 35), section 5A (examples 36 to 56) and section 6 (examples 57 to 80) of EP-A 0 392 225.

Example 3C: Analysis of Seed Lines Derived From Transformation of Tobacco with the Basic and Acidic Class III Tobacco Chitinase Genes

Binary constructions carrying the tobacco basic and acidic class III chitinases were transformed into *Nicotiana tabacum* cv *Xanthi-nc*. Leaf tissue samples were taken from T1 plants and assayed for expression of either transgene protein (by Western analysis) or transgene RNA (by Northern analysis). T1 plants found to express the transgene at high levels were advanced to T3 seed lines as described above.

Example 3D: Analysis of Transgenic Plants Expressing Class III Chitinase for Pest Resistance

Transgenic plant lines expressing one or more class III chitinase genes are assessed for resistance to numerous pests. Approximately six plants of each line are tested. Pests at the appropriate stage of their growth cycle (such as larvae) are introduced at the appropriate stage of plant development. Plants are later assayed for % of leaf or tissue area eaten, % of introduced larvae surviving, and the weight of surviving larvae.

Example 3E: Analysis of Transgenic Tobacco Plants Expressing Class III Chitinase for Pest Resistance

Transgenic tobacco plants expressing class III chitinase are assessed for resistance to numerous pests. Those tested include *Spodoptera exigua* (beet armyworm), the green peach tobacco aphid, *Manduca sexta*, and various nematodes, weevils, mites, and other pests.

Example 3F: Analysis of Transgenic Plants Expressing Basic Class III Chitinase for Resistance to *Heliothis virescens*

Transgenic tobacco lines expressing the tobacco basic class III chitinase gene were assessed for resistance to the insect *Heliothis virescens*. Eight plants of each line were tested 50 mm leaf discs were cut from the youngest leaves of transgenic plants approximately six weeks after germination and 3 larvae (2nd stage) were allowed to feed on each disc for 3-7 days. After 3-7 days the leaf discs were assessed for area eaten, % of larval survivors and weight (in mg) of larval survivors. Results of three separate experiments are shown in the tables provided below.

The results show elevated resistance to *Heliothis* in basic class III chitinase overexpressing plants when compared to non-transformed control lines *Heliothis virescens* (the tobacco budworm) causes considerable damage in tobacco crops and is particularly recalcitrant to control using pesticides as the larvae burrow deep within the plant. Furthermore, other *Heliothis* species cause similar damage to cotton and other crops.

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	(1)	%LEAF EATEN	%SURV IVORS	WT (mg) LARVAE	(2)	%LEAF EATEN	%SURV IVORS	WT(mg) LARVAE
Line								
Control		72	100	23		77	100	26
3505 C-15-5		45	92	14		65	96	19
3505 C-5-3		55	96	17		60	92	23
3505 C-4-2		57	92	19		60	100	20
3505 C-12-8		-	-	-		44	87	15

	(3)	%LEAF EATEN	%SURV IVORS	WT (mg) LARVAE
Line				

Control	65	100	53
3505 C-12-8	59	100	34

A summary of the two experiments detailed above is given below:

(1) Line:	Control	3505 C-15-5	3505 C-5-3	3505 C-4-2
Resistance Index	0	4	2.5	2

(2) Line:	Control	3505 C-15-5	3505 C-5-3	3505 C-4-2	3505 C-12-8
Resistance Index	0	2	3	3	4

(3) Line:	Control	3505 C-12-8
Resistance Index	0	3

Resistance Index:

- S = significantly more susceptible than control plants (i.e. wild-type)
- (S) = more susceptible than control plants, but not significantly in all evaluations
- 0 = like wild type plants
- 1 = some signs of resistance but not significantly different to controls
- 2 = identified resistance, significant only in some evaluations
- 3 = resistance with small significant differences in all evaluations
- 4 = high resistance, but symptoms or damage can still be observed
- 5 = total resistance. No symptoms or damage.

Example 4A: Isolation of a Genomic Clone for Arabidopsis PR-1

The *Arabidopsis* PR-1 cDNA cloned in pAPR1C-1 (sequence SEQ ID No. 17) was used as a hybridization probe in screening an *Arabidopsis* λ EMBL 3 genomic library (purchased from Clontech). Four hybridizing plaques were plaque purified using conventional techniques and λ DNA was isolated from each one with lambdasorb (Promega). The λ DNA thus isolated was digested with XhoI, electrophoresed, transferred to hybridization membrane for hybridization with the PR-1 cDNA. A fragment of 7 kb was found to hybridize to the cDNA. DNA from one of the four purified plaques was then redigested with XhoI and ligated into the XhoI site of pBluescript (Stratagene). A colony carrying a recombinant or chimaeric DNA molecule comprising the promoter region of the *Arabidopsis* PR-1 gene was identified by probing with oligonucleotide DC21 from the PR-1 coding sequence (position +110 to +84) and the plasmid contained therein was designated pAtPR1-P and deposited January 5, 1994 with the the Agricultural Research Culture Collection, International Depositing Authority, 1815 N. University Street, Peoria, Illinois 61604 under the accession no. NRRL B-21169. Restriction analysis identified the 7 kb XhoI fragment as extending 4.2 kb upstream of the ATG of the PR1 gene.

Example 4B: Fusion of the Arabidopsis PR-1 Promoter to the Firefly Luciferase Gene

Plasmid pDO432 containing a gene encoding luciferase (LUC) from firefly was received from Dr David Ow (University of California, San Diego; see Ow *et al.*, 1986; Science 234: 856). The LUC gene was excised from pDO432 by digestion with XbaI (at position +45 relative to the ATG) and SstI (approximately 1.8 kb downstream of the ATG and outside the LUC coding region). Additionally, an EcoRI-XbaI promoter fragment was

excised from pAtPR1-P; this fragment was 1.4 kb in size and delineated by an XbaI site 2.8 kb upstream of the PR-1 ATG and an EcoRI site in the pBluescript polylinker distal to the 5' end of the cloned promoter fragment (at -4.2 kb relative to the ATG). These two fragments were cloned by threeway ligation into EcoRI/SacI cleaved pBluescript thus orienting the LUC gene adjacent to the upstream PR-1 promoter fragment (pAtPR1-Q).

Subsequently, pAtPR1-P was cleaved with XbaI (at the -2.8 kb position and within the pBluescript polylinker) and religated to generate a PR-1 genomic construct without the upstream 1.4 kb promoter fragment and which thus ended 2.8 kb upstream of the PR-1 ATG (pAtPR1-R). This plasmid was used as a template in PCR with a left-to-right "topstrand" primer extending from positions -237 to -214 (DC39) upstream of the PR-1 ATG (oligo A) and a right-to-left "bottomstrand" primer comprising 15 bp of LUC coding sequence extending up to the LUC ATG and a further 19 bp of PR-1 sequence extending from the ATG into the PR-1 untranslated leader (oligo B: sequence: TTT GGC GTC TTC CAT TTT TCT AAG TTG ATA ATG G; SEQ ID No. 18). This PCR reaction was undertaken for five cycles at 94°C (30 s), 40°C (60 s), and 72°C (30 s) followed by 25 cycles at 94°C (30 s), 55°C (60 s) and 72°C (30 s) and this generated a product of 245 bp through annealing of the homologous PR-1 sequences; the fragment included a BglII site at its left end from the PR-1 promoter. A second PCR reaction was done using plasmid pDO432 as a template and using a left-to-right "topstrand" oligonucleotide which comprised 15 bp of PR-1 untranslated leader up to the PR-1 ATG and a further 12 bp of LUC sequence from the ATG into the LUC coding sequence (oligo C: sequence: TAT CAA CTT AGA AAA ATG GAA GAC GCC AAA; SEQ ID No 19) and a right-to-left "bottom strand" oligonucleotide extending from positions 332 to 312 (DC53) into the LUC coding sequence (oligo D). This PCR reaction was done under the same conditions as the one described above and generated a fragment of approximately 300 bp through annealing of the homologous LUC sequences; this fragment included a PstI site at its right end, derived from the LUC sequence amplified.

The two PCR fragments generated above were gel purified using standard procedures to remove oligonucleotides and were then themselves mixed in a further PCR reaction ("inside-outside PCR") with oligonucleotides A and D as primers. Conditions for this reaction were the same as described above. The amplified fragment was a fusion of the PR-1 promoter fragment from the first PCR reaction described above and the LUC 5' coding sequence from the second PCR reaction described above and had a BglII site at its left end and a PstI site at its right end. The fragment was gel purified and cleaved with BglII and PstI

to yield a product of 545 bp in size which was cloned into pAtPR1-R which had previously been cleaved with the same enzymes. Cleavage of the resultant plasmid (pAtPR1-S) with XbaI released a PR-1 promoter fragment extending from -2.8 kb to the XbaI site downstream of the LUC ATG, the fusion point between the PR-1 promoter and the LUC coding sequence being at the ATG. This fragment was cloned into XbaI cleaved pAtPR1-Q regenerating the full-length PR-1 promoter (4.2 kb) in operational fusion to LUC (pAtPR1-R).

Example 4C: Transfer of the *Arabidopsis* PR-1 Promoter - Firefly Luciferase Gene Fusion to pCIB200

TJS75Kan is first created by digestion of pTJS75 (Schmidhauser and Helinski, *J. Bacteriol.* 164: 446-455 (1985)) with NarI to excise the tetracycline gene, followed by insertion of an AclI fragment from pUC4K (Messing, J. and Vierra, J., *Gene* 19: 259-268 (1982)) carrying a NptII gene. pCIB 200 is then made by ligating XhoI linkers to the EcoRV fragment of pCIB7 (containing the left and right T-DNA borders, a plant selectable nos/nptII chimeric gene and the pUC polylinker, Rothstein, S.J. *et al.*, *Gene* 53: 153-161 (1987)) and cloning XhoI digested fragment into SalI digested TJS75Kan.

pAtPR1-R was cleaved with XhoI and SacI and transferred to SalI/SacI cleaved pCIB200 to create a binary vector construction (pAtPR1-S) suitable for *Arabidopsis* transformation. pAtPR1-S was then transferred to *Agrobacterium tumefaciens* strain A136/pCIB542 for transfer to *Arabidopsis* Dijon-0 using the method described by Wen-jun and Forde, *Nucl. Acids Res.* 17: 8385-8386 (1989). T2 or T3 lines carrying the PR1-LUC transgene in the homozygous state were generated for chemical induction analysis.

Example 4D: Chemical Induction of the *Arabidopsis* PR-1 Promoter

Leaves of transgenic *Arabidopsis* lines carrying the PR-1 promoter-LUC gene fusion were treated by spraying with 0.15 µg/ml INA (isonicotinic acid) for 48 h. Five days later, sprayed leaves were analyzed for luciferase activity using the Promega Luciferase Assay System (Cat. # E 1500). Transgenic lines showed 144-fold induction over controls. In other assays, luciferase activity was measured by imaging lines for bioluminescence using intensified cameras (VIM) and photon-counting image processors (ARGUS-50 or ARGUS-100) from Hamamatsu Photonic Systems (Bridgewater, NJ). Leaves were sprayed with INA and subsequently with a 5 mM solution of D-luciferin (Analytical Bioluminescence

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Laboratories, San Diego, CA) 24 h before and then immediately before imaging. Transgenic plants carrying the PR1-LUC fusion had strongly induced bioluminescence when compared to water-treated controls.

Example 5: Isolation of the Maize PR-1 and Thaumatin-like cDNAs

The maize PR-1-like cDNA PR-1mz was isolated by screening a BTH-induced cDNA library of maize with a probe matching to the PR-1 barley clone HVPR1BR (EMBL data base Accession # X74940). The DNA sequence of the gene and the deduced amino acid sequence are shown in SEQ ID NOs. 29 and 30, respectively.

The maize thaumatin-like cDNA PR-5mz was isolated by screening a BTH-induced cDNA library of maize with a probe matching to a known rice thaumatin clone (Reimann and Dudler, Plant Physiol. 101:1113-1114). The DNA sequence of the gene and the deduced amino acid sequence are shown in SEQ ID NOs. 31 and 32, respectively.

Example 6A: Transformation of Disease Resistance Genes into Maize

Ciba maize inbred line CGA00526 was transformed (Kozziel, et al., Biotechnology 11:194-200, 1993) with tobacco *PR-1a*, tobacco *SAR 8.2*, and wheat *Thionin* disease resistance genes. Constructs used for transformation include the chimeric plasmid pUBA (Toki et al, Plant Physiol. 100: 1503-1507, 1992) containing the promoter, first exon, and first intron of the maize ubiquitin gene (Ubi-1), and the coding sequence of the *bar* gene conferring resistance to phosphinothricin (Thompson et al., EMBO J. 6: 2519-2523, 1987). For cointegration purposes, the coding sequence of either the tobacco *PR1a* gene (Alexander et al, Proc. Natl. Acad. Sci. USA 90: 7327-7331, 1993), the tobacco *SAR8.2* gene (Alexander et al., 1992, Mol. Plant-Microbe Inter. 5:513-515), or the thionin gene from wheat (Rodriguez-Palenzuela et al., 1988, Gene 70:271-281), each under the control of said ubiquitin promoter were constructed into this plasmid resulting in three separate constructs for transformation. PCR techniques were used to obtain suitable restriction sites around the coding sequences of *PR1a*, *SAR8.2*, and wheat thionin, respectively. Maize tissue was bombarded with each plasmid separately or cotransformed with the *PR-1a* and *SAR8.2* constructs. Expression of the tobacco disease resistance genes *PR-1a* and *SAR 8.2*, a combination of *PR-1a* and *SAR 8.2*, and the wheat *Thionin* gene (Rodriguez-Palenzuela et al., Gene. 70:271-281, 1988) was confirmed by Northern blots.

Example 6B: Transformation of Disease Resistance Genes Into Wheat

Immature embryos of spring wheat were isolated aseptically approximately 2 weeks after pollination, and plated scutellum side uppermost on MS based medium (Murashige and Skoog, 1962 *Physiol. Plant* 15:473-439) supplemented with 2,4-D, glutamine and asparagine. Four to six hours prior to transformation embryos were transferred to medium also containing 0.25M mannitol. Constructs used for transformation included the chimeric plasmid pUBA (Toki et al, *Plant Physiol.* 100:1503-1506, 1992) containing the promoter, first exon and first intron of the maize ubiquitin gene (Ubi-1), and the coding sequence for the bar gene conferring resistance to phosphinothricin (Thomson et al, *EMBO J.* 6:2519-2523, 1987). This construct was co-transformed with a single plasmid construct containing the coding sequences for the two disease resistance genes PR1a (Alexander et al, *Proc. Natl. Acad. Sci. USA* 90:7327-7331, 1993) and the SAR 8.2 (Alexander et al, *Mol. Plant-Microbe Inter.* 6:513-515, 1992) from tobacco, expression of both sequences being driven by the double 35S promoter. For cloning purposes PCR techniques were used to obtain suitable restriction sites around the coding sequences of PR1a and SAR 8.2, respectively. Plasmid DNA was precipitated onto gold particles as described in the Dupont Biolistic manual. Embryos were bombarded using the PDS 1000 biolistic device (Weeks et al, *Plant Physiol.* 102:1977-1084, 1993) and transferred the following day to the original medium without added mannitol. Calli which developed from the bombarded embryos were selected for a morphogenic response and regenerated on medium containing Basta. Presence of transgene fragments were confirmed by PCR. Total RNA was extracted (Lagrimini et al, 1987 *Plant Physiol.* 84:438-442) and gene expression confirmed by Northern analysis. Transformed plants contained and expressed the bar gene and the PR1a gene. Some events also expressed the SAR 8.2 gene. Plants were selfed and Basta resistant progeny obtained.

Example 6C: Alternative Methods for Introducing and Expressing More Than One Anti-Pathogenic Sequence In Plant Tissue

In addition to the possibility of expressing more than one transgene in transgenic lines by the sexual crossing of lines which are transgenic for one gene, the skilled artisan recognizes that an equivalent way of generating lines transgenic for more than one gene is by the use of transformation plasmids which carry more than one gene. For example, the expression of two cDNAs in a transgenic line can be achieved by the transformation of the host plant with a vector carrying each cDNA under the independent regulation of two promoters *i.e.* the vector carries two expression cassettes in addition to sequences needed for antibiotic selection *in vitro*. Each expression cassette can also carry any signal

sequence, vacuolar targeting sequence and transcriptional terminator so desired. Vectors carrying multiple expression cassettes can be constructed for use with *Agrobacterium* transformation or direct gene transfer transformation systems.

A further method for the expression of more than one transgene in a transgenic plant line is to firstly transform with a single gene (with appropriate regulatory signals) carried on a transformation vector and subsequently transform a line selected from this transformation with a further gene (with appropriate regulatory signals) carried on a different plasmid which utilizes a different antibiotic selection system. This method is obvious to those of skill in the art.

Example 6D: Synergistic Effect of Combined Anti-Pathogenic Sequences

Overexpression of two or more PR proteins in a transgenic plant gives rise to a synergistic anti-pathogen effect. The table below shows data from an experiment in which control tobacco lines and tobacco lines rendered transgenic for PR-1a, SAR8.2, or PR-1a and SAR 8.2 were inoculated with the pathogen *Peronospora tabacina* 5 and 8 days after inoculation, percentage leaf area infected was assessed. The constructs used to obtain the transgenic tobacco lines are similar to the constructs described in examples 6A and 6B. Tobacco lines transgenic for both PR-1a and SAR8.2 were obtained according to example 6C. The left side of the table shows raw data from numerous lines of each phenotype, whereas the right side shows mean data for each phenotype. In addition to the mean values for percentage leaf area infected, the relative area compared as a percentage of the control (=100%) is presented. This enables a calculation of the expected value in the line expressing both PR-1a and SAR8.2 assuming that the individual components are additive in action. The observed values of 40.8 and 30.4 % are well below the expected values of 63.4 % ($0.756 \times 0.838 \times 100$) and 62.4 % ($0.694 \times 0.899 \times 100$) for 5 and 8 days post-inoculation. These results demonstrate that the disease resistance effects of PR-1a and SAR8.2 are synergistic in transgenic plants.

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Table A

	DAYS AFTER INOCULATION			MEAN DATA DAYS AFTER INOCULATION	
	5	8		5	8
Control (L6)	36.1	42.2	CONTROL	40.2	49.4
Control (L18)	42.8	51.1		(100)	(100)
Control (L22)	41.7	55.0			
PR-1a (L2)	35.0	42.8	PR-1a	30.4	34.3
PR-1a (L7)	25.0	25.0		(75.6)	(69.4)
PR-1a (L13)	31.1	35.0			
SAR8.2 (L1)	36.7	49.4	SAR8.2	33.7	44.7
SAR8.2 (L8)	36.1	45.0		(83.8)	(89.9)
SAR8.2 (L11)	26.7	45.0			
SAR8.2 (L21)	35.6	46.1			
SAR8.2 (L25)	33.3	37.8			
PR-1a/SAR8.2 (L17)	16.1	15.0	PR-1a/SAR8.2	16.1	15.0
				(40.8)	(30.4)
<u>Expected Additive Values:</u>				<u>63.4</u>	<u>62.4</u>

Example 7: Expression of SAR/CHX-independent genes in transgenic plants

The cDNAs described in examples 1A and 1B can be expressed in transgenic plants using techniques well known in the art. As components of the signal transduction pathway involved in SAR, the CHX-independent genes are useful for the manipulation of the SAR response. For example, the constitutive expression of key components in the SAR transduction pathway in transgenic plants will likely lead to the generation of plants with

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enhanced disease resistance characteristics and this will likely be achieved by the activation of components in the pathway downstream to the component being expressed transgenically and hence to the activation of anti-pathogenic end products. By way of illustration this may be achieved from the expression of the appropriate genes behind the constitutive 35S promoter cDNAs may be transferred to the vector pCGN1761 or pCGN1761/ENX which carry the double 35S CaMV promoter and the *tml* transcriptional terminator on a pUC-derived plasmid. Colonies carrying the cDNA in sense are recovered and the cDNA carrying expression cassette is subsequently excised and cloned into pCIB200 for use in plant transformations using *Agrobacterium*. For direct gene transfer, the cDNA-carrying expression cassette is transferred to the vector pCIB3064. Transformation to transgenic plants is undertaken using techniques well known in the art. For transformation of dicotyledonous species using binary *Agrobacterium* vectors such as pCIB200 see Alexander *et al.*, *Proc. Natl. Acad. Sci.* 90: 7327-7331 (1993), and for transformation of monocotyledonous species using direct gene transfer vectors such as pCIB3064 see Koziel *et al.*, *Biotechnology* 11: 194-200 (1993). Transgenic plants are screened for high-level expression of the appropriate cDNA by Northern or Western analysis. Plants which express high levels of the gene product are found to have enhanced resistance to plant pathogens.

Other promoters are suitable for the expression of these cDNAs in transgenic plants. These include (but are not restricted to) constitutive promoters (such as those from the ubiquitin and actin genes) and cell and tissue-specific promoters.

For genes involved in the signal transduction of SAR which may cause negative regulation of the SAR pathway, increased disease resistance can be achieved from the constitutive expression of cDNA in antisense to the gene coding sequence. The cloning and transfer of antisense sequences is undertaken in the same way as described above, except that the orientation of the cDNA is inverted to effect expression of antisense transcripts.

While the present invention has been described with reference to specific embodiments thereof, it will be appreciated that numerous variations, modifications, and embodiments are possible, and accordingly, all such variations, modifications and embodiments are to be regarded as being within the spirit and scope of the present invention.

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EP-A 249432

U.S. Patent No. 4,940,840

EP-A 392 225

DEPOSIT

NRRL B-21169

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

- (A) NAME: Ciba-Geigy AG
- (B) STREET: Klybeckstr. 141
- (C) CITY: Basle
- (E) COUNTRY: Switzerland
- (F) POSTAL CODE (ZIP): 4002
- (G) TELEPHONE: +41 61 69 11 11
- (H) TELEFAX: +41 61 696 79 76
- (I) TELEX: 962 991

(ii) TITLE OF INVENTION: CHEMICALLY REGULATABLE AND ANTI-PATHOGENIC DNA SEQUENCES AND USES THEREOF

(iii) NUMBER OF SEQUENCES: 29

(v) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.25 (EPO)

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1079 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

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CAGGTCCTTG TAGAGGTAGT GGAACCCCGA CCGGAGGGTC GGTGGGTAGC ATTGTGACAC	240
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CCAGACGTGA AATTGCTACC ATGTTTGCTC ATTTCACTCA CGAGACCGGA CATTCTGCT	420
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CATGTGCACC GGGAAAAGGC TACTTCGGTC GTGGTCCGAT CCAACTATCA TGGA ACTACA	540
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GTAGCAACCC AACTGTAGCT TTCAGGTCGG GTTTGTGGTT TTGGATGAAT AGCGTAAGGC	660
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TGATTTTCTT TTCTTATGTG ACTTCTTATG AAAGAGAATT TTAAGATTGA TAGATGTTTG	1020
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(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 952 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

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AGGTTTCTTT AACAACATTA TCAACCAAGC CGGTAATGGC TGCACGAGAA AAAGATTCAA	180
CACCGCGAC TCTTTCATTG ACGCTGCTAA TAATTTTCCA AACTTTGCAA GTTCCGTTAC	240
AAGACGTGAA ATTGCAAGCA TGTTTGCTCA TGTCCTCAC GAGACGGAC ACTTCTGCTA	300
CATAGAAGAA ATAAAGAAAA GGTACGTGG TAGGTGCGAC GAGAACGTAG AGCAGAGACC	360
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GTTTTGGATT AATAGCGTAA GCGCGTGCT GAACCAAGGA TTGGAGOCA CTATAAGAGC	540
CATCAATGGG ATGGAATGTA ACGGTGGGAA TTCAGGTGCG GTCAAGGCAA GGATTGGGTA	600
CTATAGAGAC TATTGTAGAC AGCTTGGTGT GGATCTGGT CATAACCTTA GTTGCTAAAA	660

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CGTTTACAA ACGCATACAA CGTGACGTGT CACGTGATAA TGCGGAATAA GAAGTTACAT	720
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(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1250 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

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CCAAGGAAAA CCAATGCTA ACCAGACTGT CATAGTGGAT CCAAAGCTC CGCGTGT	240
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TGACGTTCCC ACGACACCAC AACGTCTAGA GTGTGTGACC ATTGCGCATG GAGTTGTCAT	720
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GGGTGGGCOA TGCGGGGACA ACAAGGACAC GATTAACTT GGTCCATCGG AGATTGTGAC	840
AGAAGTCTCT GGAACGATTG GTGTATTTGG AGCAGCCAAT GTCAGTACA ATGCCATAAC	900
ATCACTAACC ATTACACAA ATGTCCGCAC GTACGGGCCC TTTGGAGAAC CGCAGTGTAC	960

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TCGTTTCAGT GTTCCCGTGC AGGACAAAAG CAGCATCGTG GGTTTCTTCG TGTGOGCTAG 1020
 GAAATACGTG GAGGCGCTCG GGGTTTACGT GTGTCCACCT ATTTCAAACCT AGTCCCGAAG 1080
 TGCTTCACAC TCGGTGTGCC TTCCACCTTT CTATGTTGIG CCAATAAAGT AGGTTATATA 1140
 CTTTGGTGT AATGCTATGC CCCCCCCAA GTGTGATGGT TTTCATATCA GCAAGAAGTT 1200
 TCTCTTGCTA CTATATGCAA AATAAAAAAA AAAAAAAA AAAACTCGAG 1250

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 340 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

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 AGGCAATOGA CGCAAGAGG CTCTTCATCC TAGACCACCA CGACAACTTC ATGCCTCACC 240
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(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 436 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

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(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1162 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

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GCGGCGAGGG TGGCGAAATC CAGAAGTGCT TTGTGGAATG CGTGCTGAAA AATGACTGCA	780
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(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1371 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

ATCCACCTCG ATCCAGCCAT GCTGTCTCTC AAGTCTTCTT TAGCTGTTC TCTAATCCTT	60
GTTGTCACCC TGACAGATGT GCTACOGCTG GTATCTTCTT CACGTGGCCT CGTTGGGGTA	120
CCCACTGATG GGGCACTGGA GGACTCTCTG TTGATGATGG AGAGATTCCA TGGCTGGATG	180
GCAAAGCATG GCAAGTGTGA TGCGGGAGTC GAGGAGAAAC TGCGGGGGTT TGACATATTC	240
CGCAGGAACG TAGAGTTTAT CGAGGCGGCG AACGAGATG GCAGGCTCTC GTACACCCCTC	300
GGGGTGAACC AGTTGCGCGA CTTCACCAC GAGGAGTTCC TTGCCAGCA CAACAGGCGC	360
CGTGTGGTGC CGTCAGAGGA GATGGTGATT ACAACTGCGG CTGGCGTTGT TGTCGAGGGT	420
GCCAATTGTC AGCCGGCGGC AAATGCTGTG CCTCGTAGCA TCAATTGGGT GAATCAAAGC	480
AAAGTCACCC CAGTCAAAAA TCAAGGAAAA GTATGCGGGG CTTGCTGGGC TTTTCTGOC	540
GTGGCCACGA TCGAAAGGC CTAOGCGGTC GCCAAGCGAG GCGAGCGGC GGTCTGTCC	600
GAGCAGGAGC TCATCGACTG TGACACAATC GACAGAGGCT GCACGAGCGG CGAGATGTAC	660
AATGCCTACT TCTGGGTCTT GAGGAACGGC GGCATCGCCA ACAGCTCAAC GTACCCCTAC	720
AAAGAGACTG ACGGCAAGTG CGAGAGAGGG AAAGTGCAGG AACACGCGGC CAGATCAGG	780
GACTACAAAT TGTCAAACC CAACTGCGAG GAGAAGCTCA TGGCAGCGGT GGCGGTGOGA	840
CCCGTGGCG TCGGGTTGGA CTCCAACGAC GAATGCTTCA AGTTCTACCA AGCTGGTTTG	900

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TACGACGGCA TGTGCATCAT GCACGGGGAA TACTTTGGCC CGTGCTOGTC CAACGACCGC	960
ATCCACTCCT TGGCCATTGT OGGGTACGCC GGCAAGGGGG GCGACAGGGT CAAGTACTGG	1020
ATGCCAAGA ACTCGTGGGG CGAGAAGTGG GGAAAGAAGG GCTACGTCTG GCTGAAGAAG	1080
GATGTTGATG AGCCGGAAGG CCTCTGCGGC CTTGCAATTC AGCCGGTATA TCCTATAGTC	1140
TGATCTGATC TGACGAGATC GACTGCACTG GCGGTGCATG AAAOCTACGG AAATGGCATT	1200
CACCTATATT TTGGGTTGCT CTGTATGCAT GGATGCGCCT ACTATATTTT ACTACATATA	1260
TATTCATCTC CCGCTAATAA AACTACATGT CTTGTATCC ATTTATGCAC GTTTATCCAT	1320
ATCTTGAATA AATTGGATGG ATTGGTTATC CAAAAAAAAA AAAAAAAAAA A	1371

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 723 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

CAACCGGGAA CCGGAGGCCT CCGACCCCTC AACTGCTGT GGTGACAAC ATGACCTGT	60
GCCCAAGACC TCGGGGAACG CTACCAACTC CGAACCAG TATGTCCCG CCTTGCACGA	120
GGCCGACGTC AAAGAAGGAG GTGAAGGGOG TTCTGTACCC ACGAGGOGAC GCACGTGTGT	180
CAGTGGAACG GGCAGGCCAG GTCAAACGGC GGGCTCATCG AGGGGATCG CCACTACGTG	240
CGGCTCAAGG CCGACCTGCG GCGACGCAC TGGCGCCCGC AGGGGAGOGG CGACCGTTGG	300
GACGAGGGGT ACGACGTGAC GGCCAAGTTC CTGGACTACT GCGACTCCCT CAAGGCCGGG	360
TTCTGTGCGG AGATGAACAG CAAGCTCAAG GACGGATACA GCGACACTA CTTGTGTCAG	420
ATCCTGGGGA AGAGCGTGGA CCAGCTGTGG AACGACTACA AGGCCAAGTA CCCCCAGCC	480
CCAGGGCTGA TCGACGATGC ATGCAGTTTG TTGTTGTATG TGTACCGGTC TTGTCCTACA	540
TACAGATACA TTATAGTACT TGTATTACTG TACAATTTAT GTACTGCTG GAATGGAATA	600
AATCAGCGTT GGCACGGTGT GTGTTAACGA ATTGAAGAGA CAAAGGGACC GTCTATAGGT	660
CATGTCATCG GTTGCGTGAA ATACATTGAA CATCATCACT TTCTTTACAG CAAAAAAAAA	720
AAA	723

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(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 765 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

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AAATGACAAT TGAAATTTAT TAAAGTAATA TGATCTAATA TGTTCAAACA AGCTCAATCA      60
CAATTTAATT TAATTAATAT TGACCGCGCA TCGCGCGAGT ACGACTACTA GTTCAATTTA      120
GAAAGCTGGA GCCATTGCCG GAGTAATCAT GAAATTGGCG GAGTTGACTT COGATTCCGG      180
TGATGGTGCG AGTATTCOCG AGCTTTTGAT CTCGGCCATT CCACGGGCGTG CATTTCCACC      240
TTGTTCTTCA CACAATCTGG GAAGGTATAC ACCTTCTCCA GCAGCAAGGT TGAAGTAGAG      300
ATCAATAATA TTAGGGCAAT GTTTGTAGCA CTGAGGGGAG CACAGCTTCT GGGTAAAGTG      360
GCACTCGAGG AGAGAATCAG AAGAATTCOG AAAGTATTOC TATCAACACC GCAAGCCTGA      420
ATACACTGGT CGGTTTCAAT CCAGTCTTTG AGCTTATCAG CCTCAATTTT CGACGTCTTA      480
CATGTATATA CTTCTTCACC ACTCCTTTGG AGACGTTTCT CTAACACGCA ACGCTTCCCA      540
GTAATTGAAA TCGCAAAGC GCATGAGTCC TTGTTTAGAT TCTGSCATGT TATGCTCCCT      600
AGAGTGACTT GAACACAGAA GACAAGTGCA CAAGCAACAA TAGCCAAAGT AAAGTTGTGG      660
AATGAAGCAA CCATGGTGAA AAATCTAGCA ACAATTGATC AGACTATAAA TTTTTTCTGA      720
GTATATATAG CTCCTTTTGG CTTTGGACTC TCCTTTTTTG TGGTT                      765

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(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1054 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

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GAATTCGGCA CGAGTTTGCG TATACATCAA TACTCCGGCT AAATCGGCAA ATATATCCAA	60
CAAATTAAGC CTTTTAAACC AATCAGTACG TTATCAATAT CATCAATGTC AGATOGTATT	120
GATGTCTTGG CTATGATGAT ATCAGATGAG GAATTTGCAG AGAAACTGCA GATGCAACAT	180
GTTCTCGGAG CCTCTTATGA AAGTTCCCAA GGATTCTGTG GAATTTGCAA TAAGAAAAAG	240
GGAGCAGATC GAATGTTCAA ACTCCAAATC TGCTCTOGTC ATTCTTTCTG TTCTCACTGC	300
ATAGGTCTTT ATGCCCAATC CAAGATTGCG GACAACGTTT TCCCTATAAC TTGTCCAGGT	360
TTGAGATGTC GTGTGTAAT TCAACCCGAA TCTTGTAGGT CTATCATTC AGTAAATACA	420
TTTGCAAGGT GGAAGAGGG CTCGGCOGAG TCAGCTATTC CTAATGGTGA AAAATTCTAT	480
TGTCCTTATA AAAGATGTT AGGGCTGTTC ATTCATGATC GACATGAAGA GATCGTTCAG	540
TGCATTTGTC CTCGTGCAA AAAACTGTTC TGTGCGAAGT GCGAGTCCC TTGGCACACT	600
GGGCGTGATT GTGACAAGTT TCAAAAGGTA GAAAAGACA GAGAAGATGA GCTAAACTTT	660
AAGCTGCTTG CTGAGATCAA AAAATGGAAG GAATGCCCTA AGTGCAAATC CATTGTGGAG	720
AAGGTTGATG GCTGTATACA CATGACTTGC AGGTGTAAAA TGGAAATTTG CTACGTATGT	780
GGAGGAACGT GGAGCGAAAG GCATTGGAGT TGCCAAGACT AGTTATGATA TACTATCGGA	840
TTGCCCTCTC ATACCAATCG GATGCTGAAT ATTTTGTGT CAGAATAAAA ATTATGTTGT	900
TTAGTTGGTT GTATTATGTC TTAAATTGG TGTCATGGT TTGTATTAGG AGCATTATAT	960
ATAGTACGAG TTGGTCCCTG CATAACATTT ACTTGATGTT GATTATAATA TGCTCGAGTT	1020
GGAGACCCCA AAAAAAAAAA AAAAAAACG CGAG	1054

(2) INFORMATION FOR SEQ ID NO: 11:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 760 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

CTCGTGCCGC TTAAAGTGTG CTCGAACCTC CTA CTGGTAA TGAGGATGAT GATGACCTGG	60
AATTTGAAAA TGTCCTACTGG AATGGTTCAG ATATGGCATC CGATGATACT CAAAAATCTC	120

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ATAGACCAAG GCACCGCGTA CATAAATCGT CTGGTTCCCA CAAGGCCATG AGCCGCTCOC	180
TTTCATGTGA CTOGCAATCA AAAGGATCTA TTTCTACACC TCGTGGGTCC ATGGTTGACC	240
TAAGCAAAC CTGAGATGGCT GCACTGTGGA GATATTGGCG ACACTTTAAC CTTAGGGAAG	300
CTATACCTAA CCOCTCGAAA GAGCAACTAA TTGATGTTGT OCAGAAGCAT TTCACATCTC	360
AGCAATTGGA CGAGTTGCAG GTAATTGTGG GATTGTITCA AGCTGCCAAG AGACTCAAGA	420
CAGTCTGCAA ATGACTCAAG GGAACGAACC CCTTTTGGGA ACGCCOCTAA CACTAACAAA	480
TAGTGGTACC TTGTGTCCA CTTGCTATTG GCGAGTAATA TGTGTAGTAT AATATGTATA	540
CTCCTGTTTA TCOCTTATGT CTTGCTTAGA GTTGTAGTTT ATGTAGATAA ATGAACTGCT	600
GAACTAACTC TGGAGGTCTC CTTCTGGTTG AATGTACTTC CTOGOGAGTG GGGGAAGTAC	660
TAGTACTTAA TCACATGTAT AGTCTAATGT CTTAAACTGT TAAATCATCA TTCAACCAAC	720
TGAATCTTGG TTTGGTATTA GTGACCCTCG TGCCGAATTC	760

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 396 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

TGCGCTTTA AGTGTGCTCG AACCTCCTAC TGGAATGAG GATGATGATG ACCTGGAATT	60
TGAAAATGTC CACTGGAATG GTTCAGATAT GGCATCOGAT GATACTCAA AATCTCATAG	120
ACCAAGGCAC CGCGTACATA AATCGTCTGG TTCCACAAG GNCATGAGCC GCTOCTTTTC	180
ATGTGACTCG CAATCAAAG GATCTATTTT TACACCTCGT GGGTCCATGG TTGACCTAAG	240
CAAACCTGAG ATGGCTGCAC TGTGGAGATA TTGGOGACAC TTAAOCTTA GGGAGGTATT	300
CCTAACCCCTC GAAAGAGCAC TTATTGATGT GGTCAAGCA TTCANATCTC AGAAATGGGC	360
GAGTGGAGGT ATTGTGGATT GGTCAAGCTG CAAGAG	396

(2) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 653 base pairs
- (B) TYPE: nucleic acid

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(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

CTGTTTGGT TGTTGAGTT ATTTTGCTTC TAAGAACTTT GTGAGAAATG GCTGCTAAAG	60
ATGCTACTTC ATCCGAGGAG GGACAAGTGT TOGGCTGCCA CAAGGTTGAG GAATGGAACG	120
AGTACTTCAA GAAAGGCGTT GAGACTAAGA AACTGGTGGT GGTCGATTTT ACTGCTTCAT	180
GGTGGGSCC TTGCCGTTTT ATTGCCCAAA TTCTTGCTGA CATTGCTAAG AAGATGCCCC	240
ATGTTATATT CCTCAAGGTT GATGTTGATG AACTGAAGAC TGTTCAGCG GGAATGGAGT	300
GTGGAGGCAA TGCCAACTTT TGTCTTCATT AAAGATGGAA AAGAAGTGA CAGAGTTGTT	360
GGTGCCAAGA AAGAGGAGTT GCAGCAGACC ATAGTGAAGC ATGCTGCTCC TGCTACTGTC	420
ACTGCTTGAA TCTCCTTAAT CAAGGGGATG ATATCCATA TTTAGTAGTA TTGTCTTTTG	480
TAATAACCAA GTAAC TTGTT CGAATTTTAC ACTATGGATC ACTGTATGGT TGTACTATCC	540
ACCATGTTTT TATTGCTTTT GTGAACCTTG TCTTGTGCT TGGAACTGA TTTGTGCATT	600
ACTGGTGTA GGTATATGC CCAATTCYAC AAAAAGACTA CTTTAGATT TCT	653

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1697 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

ATTCAAAATT ATTGGATCCT AGTTTTAAGT GGAACCTCTA ATTAATCAAT CATTTTCATGA	60
TCAACCTTTA CTAACATCAC CATTTTCCTT GACAACATTA TTTTCOGACT TACCATTTTC	120
CTTAACACCA TTACCTCTG GATTTTCCTT GACAATATTA CTCTCOGACT TCTCATTTTC	180
GTTCACTTTA TCAGTAATCT TGGCTTCTTC AGCAGCTGCT TCTTTCTCTT TCTCTTTCTC	240
TTTCTCAGCC CTTTCTCTT CTCTTCTCTC AGCCTTCAAT TTTGGCTCCT CCTTTCAGT	300

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TTCAAGAGCT TTAATCAGTC TCTCCAAACA AGTGTTTGCA TTTTCTTAG AAGACTTGGG	360
CATTAAATTC TCAGCAACAT CAGCAGGAGT AATATTAGTT TCCCCAATA AATGACGAAT	420
CTCAGGAAAA TGATCATGAG ACTCAATATC TAGATAATTA TTTGCAAGCA CCTTGAAGGA	480
GTCAAAGCAA CAGTATGATA ACACAATGTG TTTATCCATC CTCCTCTCC GAATTAAAGC	540
AGGGTCAAGC TTTTCCACAA AGTTGGTAGT GAAAACAATA AGCCTTTCAC CACCAATAGC	600
TGACCATAAC CCATCAATAA AGTTCAAAAG CCCAGATAAA GTCACCTCGC TTTGCTTTTT	660
CTCTCTCGA TTTTTCATCT TCTCTTGAC GGCATCTTTC TGTCTTTTA CTCTCTCTC	720
CTGTCTCTT TTTTCTCTT CCTTTGGCC GGTAAGGTCA AGCGAACAGT CGATGTCTTC	780
AATCACAATG ATAGACTTAC TAGTAGTATC TATTAATAAC TTTCTTAGCT CGGTGTTGTC	840
CTTAACCGCT GTCAATTCAA GATCATAGAC ATCATATTGT AAGAAGTTAG CCATTGCAGC	900
AATCATGCTA GACTTAACGG TTCTGGAGG ACCATATAGA AGATAACCAC GCTTCCATGC	960
CTTGCCAATC TTGGCATAAT AGTCTTTTGA CTGCTAAAT GTTGAAGGT CATCCATAAT	1020
CTCTGTGTTT TGTGTTGGCT CCATGGCTAA AGTATCAAAT GTTGATGGAT GTTCAAACAC	1080
TACTTGGCTC CACATTCTCC TCTGTATCC ATACCCACCA TCTCCTTAC TGTGTTGTTA	1140
CAACTTCTC TGCCTTCTC TTAAGTAAAT TGCCTTCCCT TGTCCAATA CATACTTCAA	1200
GTATGAAGCG GTGATAAGCT CGCGGTTCTT TCTGTGAAAC TTGAGTTTGA AATACCTCTT	1260
CTCATCTCC CTAGGGTACC AAGAAATTGT CTGTCTGCTG GCTACTTGTG GGCTAGAAAT	1320
CCACCAGACT TTCTCGCCTT TATATTCATC GGTACCTCC TCATGATCAT CCATGGTTAG	1380
TACAAGAGAT TGGCCATCTT TCACTACATT GGCTTTGAGA CGCTTAGCTT GTGTGGAGGA	1440
GTCTTGCTT AGGTACCTTT CAATTGCTAC ATAAGCTTTG CTAAGCTCGA ACCAGCATC	1500
AGTTTCATAC TCATGAAAAA TAATGTGCAT ATAAGGGTAG AATAGCTCA CGAGTTTATC	1560
GGTATACCTC CTAATATGAC CACGAAGTTC GTGAGGAAAA TAGTTCTGGT ACATGGTCCA	1620
GGCAAACATG ATTGTTGCAA TAGCTGGACC CAACTGAGTC CAAACATCTT GCATCATCAT	1680
CATCATCTAA TTTCTCT	1697

(2) INFORMATION FOR SEQ ID NO: 15

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 654 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

TTAACCTTGA GATAAGCATT AAAAAAACTC AATGGCAGGG AAGGTTGAGA AAGTGCTTGC	60
AGTACTGATG CTTGCAATGC TTCTGTTTTT GGAGCATTTC ATGGCTGCTA ATCATGAAAT	120
TAAACAACCT GAAGATAACT CTATTAGCCC TTTCTGCTTA ATAAATGTT TATATGGATG	180
CAGGGGGTTG CCACCTGCAA AAGCAGCCAT TTGTGCAGCT CAATGTTTGT TTAAGTGCGC	240
TGTCCAAGAT GAGGCCAATA TAGCTGAAAC TAAGGGCATA ATAGGTGAGA CTGCATACAA	300
CCAGTATGAT GTTGGATGTG CCTTGGCTA CTGCTCTGAG TTCCTGTTGA ATTATGATGA	360
GAGGAGGTTT AACTGCTGCA TGAATACTG TCGCGAGGGC AAAATGACCT GTCTGTGTGA	420
GGCTGCACCT TGAAGAAATG GTTGCCCTAA AATTATCGCC TCATCAAATG GAAGTACACT	480
GCTTTTTCTA CTTCCGGTGT TTAGTAGTAG TAGTAAATAA GTGAGGCATG TTACGTACTC	540
TTATGTTTTG TAATAATTAT GCTTTTAAAT AATGTAATCT GTCTGTGTGC ATACAATGCA	600
CACGACGCTA GCTACTACTT TTTATCTACT AAAAAAGAAA AGTAACTTAT TTCT	654

(2) INFORMATION FOR SEQ ID NO: 16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1031 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

GAGACAAATA CCTATAATAA GOCATTCATA ATCTTCTTGC TTCTTGTTC GAATAATGGG	60
GAATCTTTTC TGTGCGTGC TTGTGAAGCA ATCAGATGTT GCGGTCAAGG AGAGATTGG	120
CAAATTCCAA AAAGTACTTA ATCCAGGTCT CCAATTTGTT CCATGGGTCA TCGGTGATTA	180
CGTGGCGGT AACTGACCC TTGCTCTTCA GCACTCGAT GTTCAGTGTG AAACCAAAC	240
AAAGGACAAT GTGTTTGTGA CAGTGGTTGC ATCCATACAA TACAGAGTCT TAGCTGACAA	300
GGCAAGTGAT GCTTTTTACA GACTCAGCAA TCCAACCACC CAAATCAAAG CCTACGTCTT	360

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TGATGTGATC AGAGCATGTG TTCCAAAGCT GAACTTGGAC GATGTGTTTCG AGCAGAAGAA	420
TGAAATTGCC AAATCTGTGG AAGAAGAGCT AGACAAAGCC ATGACTGCTT ATGGTTACGA	480
AATCCTTCAA ACCCTAATTA TCGACATTGA GCCTGATCAA CAGGTTAAAC GTGCCATGAA	540
CGAAATCAAC GCGCGGGCGA GGATGAGAGT GGCAGCGAGC GAAAAAGCAG AGGCTGAGAA	600
AATCATTGAG ATCAAAAGAG CAGAGGGTGA AGCAGAGTCA AAGTACCTGT CGGGACTCGG	660
AATCGCTCGG CAGAGACAAG CGATCGTGGA CGGTCTTGAG AGACAGTGTG CTGGGGTTTCG	720
CAGGAAACGT GCGAGGGACG TCAGCGAAGG ATGTGTGGA CATGGTGATG ATGACTCAGT	780
ACTTTGACAC AATGAGAGAT ATCGGAGCAA CTTCTAAATC CTCTGCGGTG TTTATCCCTC	840
ACGGTCCAGG CGCGTCTCT GACGTGGCAG CGCAGATTCG AAATGGATTG TTACAGGCCA	900
ACAATGCCTC CTAATCACTC AAGTCAAATT GTCTTGGTTCG TCTCTTTATA TATTTTGGTA	960
TCTTCTTATT AAAAAGGTAA ATTTGACTTT TAATATAATG GTGTGCTTAT TGCGAAAAAA	1020
AAAAAAAAA A	1031

(2) INFORMATION FOR SEQ ID NO: 17:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 860 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

GGGAACAAAA GCTGGAGCTC CACCGGGTG GCGCGCTCT AGAACTAGTG GATCCCCCGG	60
GCTGCAGGAA TTCGGCAGCA GCAACTTAGA AAAATGAATT TTAGTGGCTA TTCTCGATT	120
TTAATCGTCT TTGTAGCTCT TGTAGGTGCT CTTGTTCTTC CCTCGAAAGC TCAAGATAGC	180
CCACAAGATT ATCTAAGGGT TCACAACCAG GCACGAGGAG CGGTAGGCGT AGGTCCCATG	240
CAGTGGGACG AGAGGGTTGC AGCCTATGCT CGGAGCTACG CAGAACAAC TAAAGGCAAC	300
TGCAGACTCA TAACTCTGG TGGGCTTAC GGGGAAACT TAGCTGGGG TAGGGTGAC	360
TTGTCTGGCG TCTCCGCCGT GAACATGTGG GTTAGCGAGA AGGCTAACTA CAACTACGCT	420
GCGAACACGT GCAATGGAGT TTGTGGTCAC TAACTCAAG TTGTTGGAG AAAGTCAGTG	480
AGACTCGGAT GTGCCAAGT GAGGTGTAAC AATGGTGGAA CCATAATCAG TTGCAACTAT	540

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GATCCTCGTG GGAATTATGT GAACGAGAAG CCATACTAAT GAAGTAATGA TGTGATCATG	600
CATACACACG TACATAAAGG ACGTGTATAT GTATCAGTAT TTCAATAAGG AGCATCATAT	660
GCAGGAYGTA TCAATATTTA TCAAATAATA CAAATAAGAG CTGAGATTAC GAGAATCTAT	720
TTAAATTAAA AGTTACATAC TTAATTATTA TAGTTATATA TGTAATAATAT GTGGCCTTTT	780
TAAAAGTTAC ATAATTAATT ATTATAGTTA ATGTCTTTCA AAAAAAAAAA AAAAAAACT	840
CGAGGGGGGG CCGGTACCC	860

(2) INFORMATION FOR SEQ ID NO: 18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 34 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: oligonucleotide primer

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

TTTGGGCTCT TCCATTTTTC TAAGTTGATA ATGG

(2) INFORMATION FOR SEQ ID NO: 19:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: oligonucleotide primer

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

TATCAACTTA GAAAAATGGA AGACGCCAAA

(2) INFORMATION FOR SEQ ID NO: 20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1064 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

AAACAATGAA CATTAAAGGTA TCATTACTTT TCATTCTACC AATATTCTTG CTTCTCCTAA	60
CCAGCAAAGT AAAAGCTGGA GATATTGTAG TCTATTGGGG CCAAGATGTA GGAGAAGGTA	120
AATTGATTGA CACATGCAAC TCTGGTCTCT ACAATATTGT CAACATTGCC TTTTATCTT	180
CTTTTGGCAA TTTCCAAACCT CCTAAACCTA ACTTAGCTGG CCATTGTGAA CCATCTTCTG	240
GTGGTTGCCA ACAGTTGACA AAAAGCATCA GACATTGTCA AAGCATAGGC ATTAAAATCA	300
TGCTCTCCAT TGGAGGTGGA ACTCCTACCT ACACATTATC CTCAGTTGAT GATGCCAGAC	360
AAGTTGCTGA TTACCTGTGG AACAATTTTC TGGGGGCCA ATCATCTTTT AGGCCACTTG	420
GAGATGCTGT ATTAGATGGC ATAGATTTTG ATATTGAACT TGGCCAACCA CATTATATTG	480
CACTTGCCAG GAGACTTTCA GAACATGGOC AACAAGGTAA AAAATTATAC TTAAGTGCAG	540
CACCACAATG TCCTTTTCCT GATAAACTTC TTAATGGTGC ATTGCAAACT GGTTTATTTG	600
ACTATGTTTG GGTCCAATTT TACAACAATC CCGAGTGCGA GTTCATGAGC AATTCAGAAA	660
ATTTCAAGAG GAGGTGGAAT CAGTGGACAT CAATCCCTGC AAAGAAGTTG TATATTGGAC	720
TTCCAGCAGC CAAGACAGCC GGGGTAATG GCTATATTCC AAAGCAAGTG CTAATGTCAC	780
AAGTTTTACC ATTTCTAAAG GGGTCTTCAA AATATGGAGG TGTCATGCTT TGAATAGAA	840
AATTTGATGT CCAATGTGGC TATAGCTCTG CTATCAGGGG TGCTGTTTAA GTTCTGAATG	900
AACAAGGCGC CCTGAATCG CTATAAGCCA TCGTTAAGGC CTAAATAAAG CAAGTTAATT	960
TGCTGTTATC TGCTAGAAA GTACTTAAGT TTTAATTTGT ACTGATGAAA ATGTGAAGGT	1020
CATCTTGTTT CCTTCTTGAT AATAGTAGTA CTATGGTTCT CTTT	1064

(2) INFORMATION FOR SEQ ID NO: 21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1018 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

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TGAAGTGTTA GCACACACAC ACAAAAAAAAA CTTAAAGTTA TGATCAAATA TAGTTTTCTT	60
TTGACAGCAT TAGTGCTATT TCTTCGAGCA TTAAACTAG AAGCAGGGGA TATAGTAATA	120
TATTGGGGCC AAAATGGGAA TGAAGGTAGC TTAGCTGACA CTTGTGCAAC AAATAACTAT	180
GCCATTGTCA ATATTGCTTT CCTGTAGTT TTTGGGAATG GCCAAAATCC AGTGCTAAAT	240
TTAGCTGGTC ATTGTGATCC AAATGCTGGT GCATGCACTG GCTTAAGCAA TGACATTAGA	300
GCTTGTCAA ACCAAGGCAT CAAAGTTATG CTTTCTCTTG GTGGTGGTGC TGAAGCTAT	360
TTTCTTTCTT CTGCTGATGA TGCTAGGAAT GTGGCAAATT AATTGTGGAA CAATTATCTT	420
GGAGGTCAAT CAAACACACG TCCACTAGGA GATGCAGTTC TAGATGGAAT TGATTTTGAT	480
ATAGAAGGCG GGACAACACA ACATTGGGAT GAATTAGCAA AACTCTATC ACAATTTAGC	540
CAACAAAGGA AAGTATACTT AACTGCAGCT CCACAATGTC CATTOCCAGA TACATGGTTA	600
AATGGGGCAC TTCCACTGG CTTATTTGAT TATGTTTGGG TTCAATTTTA CAATAATCCA	660
CCGTGTCAAT ACTCCGGTGG GAGCGCGGAC AATTTAAAAA ATTACTGGAA TCAGTGGAAC	720
GCGATTCAAG CTGGAAAAAT TTTTCTGGGA TTGCCAGCAG CTCAGGAGC AGCTGGAAGT	780
GGTTTTATAC CATCTGATGT TCTTGTCTCT CAGGTTTAC CATTAAATTA TGGTTCACCA	840
AAGTATGGGG GTGTTATGCT TTGGTCTAAA TTTTATGACA ATGGTTATAG CTCTGCTATT	900
AAGGCTAATG TTTGAGATAT ATGATCATAG CTAGTCAGCT TGTATTAATA TGATGACGTC	960
AATAATGTTA TATTATAAAC TATATAGTAC TCAATAATAA GGCTTTGAAA GTTACTTA	1018

(2) INFORMATION FOR SEQ ID NO: 22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 12 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

GGGATCCCTG CA

12

(2) INFORMATION FOR SEQ ID NO: 23:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 12 base pairs

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- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

ATAGTCTTGT TGAGAGTT

18

(2) INFORMATION FOR SEQ ID NO: 24:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

TCACGGGTTG GGGTTTCTAC

20

(2) INFORMATION FOR SEQ ID NO: 25:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 18 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 25:

AGGAGATGGT TTGGTGGA

18

(2) INFORMATION FOR SEQ ID NO: 26:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26:

ATACGTTCTA CTATCATAGT

51

(2) INFORMATION FOR SEQ ID NO: 27:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:

His Phe Cys Tyr Ile Glu Glu
1 5

(2) INFORMATION FOR SEQ ID NO: 28:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 6 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:

Ile Arg Ala Ile Asn Gly
1 5

(2) INFORMATION FOR SEQ ID NO: 29:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 731 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 40..528

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(D) OTHER INFORMATION:/note= "PR-1mz cDNA clone"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:

GAATTCGGG AACTAGTAAT TATCAGCAAA CAACAAACAA TGGCACOGAG GCTAGCGTGC	60
CTCCTAGCTC TGGCCATGGC AGCCATCGTC GTGGCGCCGT GCACGGCCCA GAACTOGCCG	120
CAGGACTACG TGGACCCGCA CAACGCGGCG CGGCGGACG TGGGCGTOGG GCGGTGTCC	180
TGGGACGACA CCGTGGCOGC GTACGCGCAG AGCTACGCGG CGCAGGCCA GGGGACTGC	240
AAGCTGATCC ACTCOGGGGG GOCCTACGGC GAGAACCTCT TCTGGGGCTC CGCOGGGCGC	300
GACTGGTGGG CGTCCGACGC CGTGGGCTCC TGGGTGTCCG AGAAGCAGTA CTACGACCAC	360
GACACCAACA GCTGCGCGGA GGGGCAGGTG TGCGGOCCT ACACGCAGGT GGTGTGGCGC	420
GACTCCACOG CCATCGGCTG TGCCCGGTC GTCTGCGACA ACAACGCCGG CGTCTTCATC	480
ATCTGCAGCT ACAACCGGCC GGGCAACGTC GTGGGCGAGA GCGCTACTA GACTGTCATG	540
CATACTACAA TTATATATAT ATACGCTTAA TATTAACTTC AGCATGCATG CATATTATAA	600
ATAGTGTGTG CAACTCTGTA TATCATTATA TTACGATGAT TATATTGTTG ATGAACATAA	660
TAAGTCAATA AAACCATGCA GGCGATTTGT GAGCCAAAA AAAAAAAAAA AAAAAAAAAA	720
AAAAACTCGA G	731

(2) INFORMATION FOR SEQ ID NO: 30:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 163 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 30:

Met	Ala	Pro	Arg	Leu	Ala	Cys	Leu	Leu	Ala	Leu	Ala	Met	Ala	Ala	Ile
1				5				10					15		
Val	Val	Ala	Pro	Cys	Thr	Ala	Gln	Asn	Ser	Pro	Gln	Asp	Tyr	Val	Asp
			20					25					30		
Pro	His	Asn	Ala	Ala	Arg	Ala	Asp	Val	Gly	Val	Gly	Pro	Val	Ser	Trp
		35					40					45			
Asp	Asp	Thr	Val	Ala	Ala	Tyr	Ala	Gln	Ser	Tyr	Ala	Ala	Gln	Arg	Gln
	50					55					60				
Gly	Asp	Cys	Lys	Leu	Ile	His	Ser	Gly	Gly	Pro	Tyr	Gly	Glu	Asn	Leu

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65		70		75		80									
Phe	Trp	Gly	Ser	Ala	Gly	Ala	Asp	Trp	Ser	Ala	Ser	Asp	Ala	Val	Gly
				85					90					95	
Ser	Trp	Val	Ser	Glu	Lys	Gln	Tyr	Tyr	Asp	His	Asp	Thr	Asn	Ser	Cys
		100						105					110		
Ala	Glu	Gly	Gln	Val	Cys	Gly	His	Tyr	Thr	Gln	Val	Val	Trp	Arg	Asp
		115						120					125		
Ser	Thr	Ala	Ile	Gly	Cys	Ala	Arg	Val	Val	Cys	Asp	Asn	Asn	Ala	Gly
	130					135					140				
Val	Phe	Ile	Ile	Cys	Ser	Tyr	Asn	Pro	Pro	Gly	Asn	Val	Val	Gly	Glu
145				150						155					160
Ser	Pro	Tyr													

(2) INFORMATION FOR SEQ ID NO: 31:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 831 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 72..587
- (D) OTHER INFORMATION: /note= "PR-5mz cDNA clone"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 31:

GAATTCCGGA	GOCTAGCTCT	ATAGCTCGAG	TATTGCTTGC	TCTCCACAAG	AAACAAGGCA	60
GAGCAOCCAA	AATGGCGGCC	GOGTCTCGG	TCCTCTGCT	GCTCTGGOC	GOOGCTTGG	120
CTGGCATGAG	CGCCAACGCC	GCAOCTTCA	CCATCACCAA	CAACTGCGGG	TTACCGTGT	180
GGCCCGCGGC	CACCCCGGTG	GGCGGGGCA	CGCAGCTGAA	COCGGCGGG	ACGTGGACCG	240
TCAACGTGCC	GGCGGCACC	AGCTCCGGC	GCGTGTGGG	CGCAOCCGC	TGCTCCTTCA	300
ACGGCAACAG	CGGGAGCTGC	CAGAOGGOG	ACTGCGGGG	CGGCTGGOC	TGCAOCTGT	360
CGGGGCAGOC	GCGCTGACG	CTGGCGAGT	TCACCATCGG	CGGCAGOCAG	GACTTCTACG	420

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ACATCTCGGT CATCGACGGC TACAACCTOG CCATGGCCTT CTCCTGCAGC ACOGGCGTGC 480
 GGCTGGTGTG CACGGACCCT GGATGCCCCG ACGCGTACCA CAACCCCCC GACATGAAGA 540
 CCCATGCCTG TGGCGGCAAC AGCAACTACC AAGTCACCTT CTGCCCCTGA TCGGCCGGAA 600
 TAGGCTCTGC ATGCATGGGC TAGTGATTG GATTTTGCAG GAATAATTCT GATTGGATT 660
 TGCACAATAA TATATAAGCG TGCATTACAC GTGCACGCAT TTGTATGTAC GCGGTATAC 720
 GTCGTACCTA CGTAGCTGGA TTGTGTGCG TGCTTGTAC CAAATATTAA TAAGAATAAA 780
 CAAATATGAC TCGTGTGATA TATATATAAA AAAAAAAAAA AAAAATOCA G 831

(2) INFORMATION FOR SEQ ID NO: 32:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 172 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 32:

Met Ala Ala Ala Ser Ser Val Leu Leu Leu Leu Leu Ala Ala Ala Leu
 1 5 10 15
 Ala Gly Met Ser Ala Asn Ala Ala Thr Phe Thr Ile Thr Asn Asn Cys
 20 25 30
 Gly Phe Thr Val Trp Pro Ala Ala Thr Pro Val Gly Gly Gly Thr Gln
 35 40 45
 Leu Asn Pro Gly Gly Thr Trp Thr Val Asn Val Pro Ala Gly Thr Ser
 50 55 60
 Ser Gly Arg Val Trp Gly Arg Thr Gly Cys Ser Phe Asn Gly Asn Ser
 65 70 75 80
 Gly Ser Cys Gln Thr Gly Asp Cys Gly Gly Ala Leu Ala Cys Thr Leu
 85 90 95
 Ser Gly Gln Pro Pro Leu Thr Leu Ala Glu Phe Thr Ile Gly Gly Ser
 100 105 110
 Gln Asp Phe Tyr Asp Ile Ser Val Ile Asp Gly Tyr Asn Leu Ala Met
 115 120 125
 Ala Phe Ser Cys Ser Thr Gly Val Arg Leu Val Cys Thr Asp Pro Gly
 130 135 140
 Cys Pro Asp Ala Tyr His Asn Pro Pro Asp Met Lys Thr His Ala Cys
 145 150 155 160

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Gly Gly Asn Ser Asn Tyr Gln Val Thr Phe Cys Pro
 165 170

(2) INFORMATION FOR SEQ ID NO: 33:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 790 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 1..790
- (D) OTHER INFORMATION: /note= "cDNA called PSI-1"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 33:

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GGCAGAGGT CATATCGTT GAAAACATCA TTCACTCTTC TCTGTGTC TCCTTTTCAC      60
CCGAAATTGT CTCTGACACC GNCTTGNINC AGNCATGGCT AATGCTGCGT CAGGAATGGC    120
AGTCCATGAT GACTGCWWRY KARKAKYYCY KGGAMIGAAG GCGAAAAGGA CACAMCRYTT    180
CATTGTCTAC AAGATTGAGG AGAAGCAGAA GCAAGTGATT GTTGAGAAAG TTGGTGAACC    240
TATTCTAACT TACGAGGACT TTGCAGCAAG TCTTCCAGCT GACGAATGMC GATACGCCAT    300
YTATGATTTC GACTTTGTCA CTGCAGAGAA TTGCCAGAAG AGCAAGATTT TCTTCATTGC    360
ATGGTGTCCC GACGTAGCAA AGGTGAGAAG CAAGATGATC TATGOGAGCT CTAAGGACAG    420
GTTCAAGCGT GAACTTGATG GAATTCAAGT GGAGCTTCAA GCAACTGATC CAACTGAGAT    480
GGATCTTGAT GTTTTGAAAA GCOGCGTCAA CTAAAAGAAA CTCTCTTTGA ATAAGCTTCC    540
GATTATATTC GACTATTATG AAATGGTGTT TTCTATTCTC TTACAGTCTC TTGTGACTGT    600
TAGATACTAC TGACACCCCTT CATTCTATCT GTCTGGTCTG TTCTCTGTGT CTATGTTACC    660
TTACAGTGTC TCATCTATAT TCAAAATTAT CACAAGGTTT GTGCTGTGTT TGCTTCCTTT    720
GTTTCAACTC TACCAAAGCT TAAGATTAAG ATTTGGGTAA GATGATTCTT TTAAAAA      780
AAAAA      790

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(2) INFORMATION FOR SEQ ID NO: 34:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 508 base pairs
- (B) TYPE: nucleic acid

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- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION:1..508
- (D) OTHER INFORMATION:/note= "cDNA called PSI-2"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 34:

CCTCCAAGAA CCTAGTGGGA TCCCCCOGGC CTGCAGGAAT TOGGCACCGA GCTCCTTGCC	60
GAAATTCGGC ACGAGTGTG ATCTAAATCC TCACIAGGTG GTTOOGTTTA AGCTTAGCAA	120
CGCCAAAGAT CTAGACAATA ATTGCAAGTT ACCGGGTTGA TCCTATCTTC CGTCCCGAAG	180
CCGTCCCTGA TCCAGCCCAA GATGTTTTGT GGTCAGATGA CAAAGTAGAT CCACGGGTAT	240
ATATTATGGT AAAGTTGGCG GCOGAAGGAT TGAAGTTCAA TAATGATATG TTTTCTGGTG	300
GTTGCATCCC ATCTGAAGTG GTTATAGCCC CCAAAAACAA AAAATCTCTA CAATTTTGAA	360
AGGTGTTGCT GGCAGGAAAT CTGCTAAGCA AGGAAGGATC TOCAAGAATC CAACTGGGAA	420
TAGAAGGATC CATCAATCAT TCTCTAATGA GGAGGAATCT GTTAACTTT GTTATTGACT	480
AATAATCCTA TTATCGOCTA AAGCTGTT	508

(2) INFORMATION FOR SEQ ID NO: 35:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 570 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION:1..570
- (D) OTHER INFORMATION:/note= "cDNA called PSI-3"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 35:

GGCAGGAGCT CGTGCGCTC GTGCCGAATT OGGCAGGAG CAACATTTC AACACCATCG	60
ACGGTGATAG GCCTCGGAGG ATCATCCATC ACCACCAAAC CCTTCTCTTC ATCCTTTTTA	120

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AAACCAACAT TAAGCGCCAA GAACCCTTTG AGACTOGCCG GTGCATCGGG AGGAAGAGTC	180
ACTTGCTTTG AGAGGAACTG GTTGAGGAGA GATTGAACG TGGTAGGATT TGGGCTGATC	240
GGATGGCTAG CTCGCTOGAG CATTCCAGOG ATAAATGGGA AGAGCCTGAC GGGTCTCTTC	300
TTCGATAGCA TCGGAACTGA GCTCGCTCAC TTCCCGACTC CTCAGCTCT CACTTCACAG	360
TTCTGGTTGT GGTGGTTAC GTGGCACITA GGCTCTTCC TCTGCCTCAC TTTCGGACAA	420
ATCGGATTCA AGGGCAGGAC TGAAGATTAC TTCTAAGGAT AACTATTCTT GTTTTCGTTT	480
GTAATATATG CTCTCTCTGG KTATGTGTAA TATTATCAAT CAAACAAAG CTTTCTTTGC	540
TATTTGATGT TTCATTCTAT AAAAAAAAAA	570

(2) INFORMATION FOR SEQ ID NO: 36:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1418 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: misc feature
- (B) LOCATION: 1..1418
- (D) OTHER INFORMATION: /note= "cDNA called PSI-4"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 36:

GGCAGAGCA GAATCCTTAG CATCAGAGAA TTCTTCAATG ACAGACACTT ATAACCAGCA	60
GAGAGGTCTT GTCAACCAAC TAAAAGATGA CATGGAGAGG CTATATCAAC AGATCCAAGC	120
CCAAATGGGC GAACTTGAGT CTGTCAGGGT TGAGTATGCA AACGCGCAGC TAGAATGTAA	180
TGCTGCTGAT GAGCGTTCCG AAATACTGGC TTCTGAAGTC ATCAGTTTGG AAGATAAGGC	240
TCTCAGACTC AGGATCTAAT GAGTTAAAGC TGGAGAGGGA ACTGGAGAAG GCACAAAACA	300
GAAATTGTTA TCTTACAAGA AAAAATTGCA GGAGCTTAGA GAAAGGATCC GTCAAAGATT	360
TACAAATCTT ACTATTAAAG CTCTTCAGGA AAGAGAAAGG AAGGGTCOCT ACAAACCTAT	420
GGGTGCAGGA AAGCTTCATC TGGTGGGGAA AAATCCACTG GATCTTTGAG TTAAAACTCA	480
ACTAGCAGAA AGAACGTGTC AACCTCTACA GAAGGTCCTG CAATCTCAGA TACTACGCCA	540
GAGAGCTOCA ACCAGGAAAC AGATTCTACT ACTCTGCTCG AAAGTGATTC ATCTWATACA	600

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GCTATCATTC CTGAAACTAG ACAATTAAC TTTGAAGGCT TTTCATTGAG CGTCCCAGCT	660
GATCAGATGA GAGTGATTCA TAACATTAAT ACGCTGATTG CTGAGTTGGC AATTGAGAAG	720
GAAGAACTGG TGCAAGCACT GTCATCTGAG TTATCTOGAA GTGCGCATGT GCAGGAGCTG	780
AACAAAGAGT TATCCAGAAA ACTTGAAGCG CAGACGCAAA GGTTAGAGCT TGTAACAGCG	840
CAGAAGATGG CCATAGACAA TGTTTCACCA GAAAAGCAGC AGSCGGACAC TCATGTTGTT	900
CAAGAGAGAA CMCCGATTGC AGATGAAGGC GATGAGGTGG TAGAAAGGGT TCTWAGGATG	960
GATCATGAAG ATGTTCCAG GAGGACOGTC GAAAAGAAGG ACAAGCAAGC TTCTCTAATG	1020
CATGGATGGC TTGCTCAGTC GTGACCOCTG TTGCCATTCT GTATATCCCG GGCACAGCCA	1080
TGGATTGCTT CTTCTCCTTT AGKKKTGGAA GAATTCATTT GGTTCCTTTG ATCCCCAAAT	1140
TTTGGATTTT ATTGGATTGA GOMCAAATGG TATAGTGAGA AACACCATGA ACTTGAAGAG	1200
TTCCAAATGG ATAGAGGAGA CTCGGAAGCT TAAACAGAGA GGAAGGTAGT TGTATATGA	1260
GTGCGTTTTT GTCTGAATTT TGTAATTGTT GAATATATAG CCAAGTTAAA GTGTTCTCTC	1320
TAGCAAATAT AATTTACCAA GAAAAAATCT TGTATGGTTT CGGATATATT TTATCTTCTC	1380
GTTTTAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAAAA	1418

(2) INFORMATION FOR SEQ ID NO: 37:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 913 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1..913
- (D) OTHER INFORMATION: /note= "cDNA called PSI-5"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 37:

GGCAGGAGAG ACAATCAGAT TCTTTACTCT TACAACGGTG GAGACCAAAC CAACGAGAGT	60
CTTGCTGCTC TGTGTTTAGA AAAGTCTCCT CCTTTTCATA CTTGTTACTT TGAAACTATT	120
GGTAAAAGAA GATTTGGGTT TTTAATTGGA GATGGGTTTG TTTATTTGCG CATTGTTGAT	180
GAGGTTTGA AGAGATCTAG TGTTCTTAAG TTCTTGAAC ATTTGGAGAG ATGAATTTAA	240

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GAAAGCTGCT AGGGAGAATT CTAGAGGAAG TTTCAC TGCT ATGATTGGTT CTATTAATGT	300
TGAAGATCAG CTTGTTCC TG TTGTTACTAG ACTTATAGCT TCTCTTGAAC GTGTTGCTGA	360
GAGTAGTAGT AACAAATGAAT TGAAGAGTAG TAATCTTGGT GAACAAAGCG AAGGTTTCGAA	420
TTCTACTAAA GCTCCGTTGT TGGGGAGGTT AAGTAAGCAA GAGAAGAAGA AAGGGAAAGA	480
TCATGTGATT GAGCTTGAGG AACACAGGAA GTCTAATGAT AGAGGGAACA TAACCGATGA	540
TTCAGCGGGA GCTGGAACGT CGTTGGAGAA GGAGTGTGTG TCGAGTAGTG GACGTTCTGT	600
TACTCAAAGC TTTGAATGGA AATGGCGGCG GYTAGTTCAG ATTGTTCTTG CTATTGATGC	660
AGCTATTTGC TTGACACTGT TTGGTATTTG GCTGGCTATA TGTCGGGGTA TCGAGTGTAC	720
ACGTTTCGTGA TTGTGCAGAC AGAAAATCGG ATAAACAGAG TCGATTCTTT ACATTCCCTT	780
GTGCATTCCA ATAGCCAAAA AATATGTTGA TTGTAATGTG AACCATTCTT AAGTTATAAT	840
TTGCCTTATG TATTGCTTTA GTCACAGCTT TTGAAATATG TACAGTCGGA TTCTAGAAAA	900
AAAAAAAAAA AAA	913

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WHAT IS CLAIMED IS:

1. A recombinant or chimaeric DNA molecule comprising a plant SAR gene different from the DNA molecule p1.4.3 (SEQ ID NO. 12 / SEQ ID NO. 13), wherein the wild-type gene corresponding to said SAR gene can be chemically induced in a plant in a protein-synthesis independent manner.
2. A DNA molecule according to claim 1, wherein the corresponding wild-type gene can be chemically induced in a plant despite the presence of the protein synthesis inhibitor cycloheximide.
3. A DNA molecule according to claim 1, wherein the corresponding wild-type gene can be induced in a plant by salicylic acid, isonicotinic acid or methyl benzo-1,2,3-thiadiazole-7-carboxylate.
4. A cDNA molecule according to claim 1.
5. A DNA molecule according to claim 1 comprising DNA selected from the group of DNA molecules p11.30.13 (SEQ ID NO. 11), p66B1 (SEQ ID NO. 14), pDPA2 (SEQ ID NO. 16), PSI-1 (SEQ ID NO. 34), PSI-2 (SEQ ID NO. 35), PSI-3 (SEQ ID NO. 36), PSI-4 (SEQ ID NO. 37), and PSI-5 (SEQ ID NO. 38).
6. A recombinant or chimaeric DNA molecule comprising a plant SAR gene different from the DNA molecule p1.4.3 (SEQ ID NO. 12 / SEQ ID NO. 13) obtainable by a method comprising
 - (a) cloning genes which are chemically inducible by differential screening of cDNA libraries;
 - (b) further analyzing the clones obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor; and
 - (c) identifying clones which hybridise with both RNA chemically induced in the absence of a protein synthesis inhibitor and RNA chemically induced in the presence of a protein synthesis inhibitor.

7. A recombinant or chimaeric DNA molecule comprising a plant SAR gene different from the DNA molecule p1.4.3 (SEQ ID NO. 12 / SEQ ID NO. 13) obtainable by a method comprising
 - (a) cloning genes which are chemically inducible in the presence or absence of a protein synthesis inhibitor by differential display of isolated RNA using polymerase chain reaction;
 - (b) further analyzing the amplification products obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor; and
 - (c) identifying amplification products which hybridise with both RNA chemically induced in the absence of a protein synthesis inhibitor and RNA chemically induced in the presence of a protein synthesis inhibitor.
8. A recombinant or chimaeric DNA molecule comprising a plant SAR gene, wherein the wild-type gene corresponding to said SAR gene can be chemically induced in a plant in a protein-synthesis dependent manner.
9. A DNA molecule according to claim 8 wherein the corresponding wild-type gene can only be chemically induced in the absence of the protein synthesis inhibitor cycloheximide.
10. A DNA molecule according to claim 8 wherein the corresponding wild-type gene can be induced by salicylic acid, isonicotinic acid or methyl benzo-1,2,3-thiadiazole-7-carboxylate.
11. A cDNA molecule according to claims 8.
12. A DNA molecule according to claim 8 comprising DNA selected from the group of DNA molecules p1.1.1 (SEQ ID NO. 9), p.11.31.4 (SEQ ID NO. 10), and p.14.22.3 (SEQ ID NO. 15).
13. A recombinant or chimaeric DNA molecule comprising a plant SAR gene obtainable by a method comprising
 - (a) cloning genes which are chemically inducible by differential screening of cDNA libraries;

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- (b) further analyzing the clones obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor; and
 - (c) identifying clones which hybridise with RNA chemically induced in the absence of a protein synthesis inhibitor but not with RNA chemically induced in the presence of a protein synthesis inhibitor.
14. A recombinant or chimaeric DNA molecule comprising a plant SAR gene obtainable by a method comprising
- (a) cloning genes which are chemically inducible in the absence of a protein synthesis inhibitor by differential display of isolated RNA using polymerase chain reaction;
 - (b) further analyzing the amplification products obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor; and
 - (c) identifying amplification products which hybridise with RNA chemically induced in the absence of a protein synthesis inhibitor but not with RNA chemically induced in the presence of a protein synthesis inhibitor.
15. A recombinant or chimaeric DNA molecule comprising the cDNA of a wild-type wheat gene which wild-type gene can be chemically induced in a plant.
16. A DNA molecule according to claim 15 wherein the corresponding wild-type gene can be induced in a plant by methyl benzo-1,2,3-thiadiazole-7-carboxylate.
17. A DNA molecule according to claim 15 comprising DNA selected from the group of DNA molecules WCI-1 (SEQ ID NO. 3), WCI-2 (SEQ ID NO. 4 / SEQ ID NO. 5), WCI-3 (SEQ ID NO. 6), WCI-4 (SEQ ID NO. 7), and WCI-5 (SEQ ID NO. 8).
18. A recombinant or chimaeric DNA molecule encoding Arabidopsis class IV chitinase, Tobacco chitinase/lysozyme, Maize PR-1, or Maize PR-5.
19. A DNA molecule according to claim 18 encoding Arabidopsis class IV-chitinase-type A or B.

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20. A DNA molecule according to claim 18 comprising the DNA sequence pChit4-TA (SEQ ID NO. 1), pChit4-TB (SEQ ID NO. 2), pBSCL2 (SEQ ID NO. 20), or pBSTCL226 (SEQ ID NO. 21).
21. A DNA molecule according to claim 18 comprising the DNA sequence PR-1mz (SEQ ID NO. 30), or PR-5mz (SEQ ID NO. 32).
22. A recombinant or chimaeric DNA molecule comprising the promoter region of the Arabidopsis PR-1 gene.
23. A DNA molecule according to claim 18 comprising 4.2 kb upstream of the start codon of the PR-1 wild-type gene.
24. A method for obtaining a recombinant or chimaeric DNA molecule encoding a gene which corresponds to a wild-type gene which can be chemically induced in a protein-synthesis independent manner, comprising
 - (a) cloning genes which are chemically inducible by differential screening of cDNA libraries;
 - (b) further analyzing the clones obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor; and
 - (c) identifying clones which hybridise with both RNA chemically induced in the absence of a protein synthesis inhibitor and RNA chemically induced in the presence of a protein synthesis inhibitor.
25. A method for obtaining a recombinant or chimaeric DNA molecule encoding a gene which corresponds to a wild-type gene which can be chemically induced in a protein-synthesis independent manner, comprising
 - (a) cloning genes which are chemically inducible in the presence or absence of a protein synthesis inhibitor by differential display of isolated RNA using polymerase chain reaction;
 - (b) further analyzing the amplification products obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor; and

- (c) identifying amplification products which hybridise with both RNA chemically induced in the absence of a protein synthesis inhibitor and RNA chemically induced in the presence of a protein synthesis inhibitor.
26. A method according to claims 24 or 25, wherein cycloheximide is used as the protein synthesis inhibitor.
27. A method for obtaining a recombinant or chimaeric DNA molecule encoding a gene which corresponds to a wild-type gene which can be chemically induced in a protein-synthesis dependent manner, comprising
- (a) cloning genes which are chemically inducible by differential screening of cDNA libraries;
 - (b) further analyzing the clones obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor; and
 - (c) identifying clones which hybridise with RNA chemically induced in the absence of a protein synthesis inhibitor but not with RNA chemically induced in the presence of a protein synthesis inhibitor.
28. A method for obtaining a recombinant or chimaeric DNA molecule encoding a gene which corresponds to a wild-type gene which can be chemically induced in a protein-synthesis dependent manner, comprising
- (a) cloning genes which are chemically inducible in the absence of a protein synthesis inhibitor by differential display of isolated RNA using polymerase chain reaction;
 - (b) further analyzing the amplification products obtained in step (a) by Northern hybridisation to RNA isolated from cells chemically induced in the presence or absence of a protein synthesis inhibitor; and
 - (c) identifying amplification products which hybridise with RNA chemically induced in the absence of a protein synthesis inhibitor but not with RNA chemically induced in the presence of a protein synthesis inhibitor.
29. A method according to claims 27 or 28, wherein cycloheximide is used as the protein synthesis inhibitor.
30. A method for obtaining a recombinant or chimaeric DNA molecule comprising the cDNA of a wild-type wheat gene which wild-type gene can be chemically induced in a wheat plant, comprising

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- (a) cloning genes which are chemically inducible by differential screening of wheat cDNA libraries;
 - (b) further analyzing the clones obtained in step (a) by Northern hybridisation to RNA isolated from wheat cells chemically induced; and
 - (c) identifying amplification products which hybridise with both RNA chemically induced in the absence of a protein synthesis inhibitor and RNA chemically induced in the presence of a protein synthesis inhibitor.
31. A method for obtaining a recombinant or chimaeric DNA molecule encoding plant class IV chitinase comprising amplifying by the polymerase chain reaction DNA molecules from plant DNA using 5' and 3' oligonucleotides designed degenerate for the peptides HFCYIEE (SEQ ID NO. 27) and IRAING (SEQ ID NO. 28), respectively.
32. A method for obtaining a recombinant or chimaeric DNA molecule encoding plant chitinase/lysozyme comprising screening a plant cDNA library with a cDNA probe encoding cucumber chitinase/lysozyme.
33. A method for obtaining a recombinant or chimaeric DNA molecule comprising the promoter region of the Arabidopsis PR-1 gene comprising
- (a) screening of an Arabidopsis cDNA library with a probe of Arabidopsis PR-1 c-DNA; and
 - (b) identifying a clone carrying the promoter fragment by hybridisation to an oligonucleotide corresponding to coding sequences of the Arabidopsis PR-1 gene.
34. A method of improving protection of a plant against a pest comprising transgenically expressing in said plant two or more DNA molecules encoding anti-pathogenic proteins, wherein the transgenically expressed proteins exert a synergistic effect.
35. A method according to claim 34, wherein two or more DNA molecules according to any one of claims 1-19 are transgenically expressed.
36. A method of protecting plants against pests comprising transgenically expressing in said plant class III chitinase.
37. A method according to claim 36, wherein tobacco plant chitinase III is transgenically expressed.

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38. A transgenic plant and the progeny thereof comprising a DNA molecule according to any one of claims 1 to 23 displaying anti-pathogenic properties.
39. A transgenic plant according to claim 38 expressing transgenic Tobacco chitinase/lysozyme.
40. A transgenic plant and the progeny thereof comprising two or more DNA molecules encoding anti-pathogenic proteins, which plant displays synergistic anti-pathogenic properties.
41. A transgenic plant according to claim 40 comprising two or more DNA molecules according to any one of claims 1-23.
42. Seed of a transgenic plant according to any one of claims 38-41.
43. Use of a DNA molecule according to any one of claims 1 to 7 to achieve expression of a protein-synthesis independent plant SAR gene in transgenic plants.
44. Use of a DNA molecule according to any one of claims 8 to 14 to achieve expression of a protein-synthesis dependent plant SAR gene in transgenic plants.
45. Use of a recombinant or chimaeric DNA molecule according to any one of claims 18 to 21 to obtain transgenic plants displaying anti-pathogenic properties.
46. Use of the promoter region according to claim 22 to chemically induce a gene in operable linkage with said promoter.